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(51) International Patent Classification ⁵ : C12P 19/34, 21/06, C12N 15/00 A61K 35/14, 37/00, 39/00 C07K 13/00, C07H 21/04	A1	(11) International Publication Number: WO 94/00590 (43) International Publication Date: 6 January 1994 (06.01.94)
(21) International Application Number: PCT/US93/05965 (22) International Filing Date: 22 June 1993 (22.06.93) (30) Priority data: 904,072 23 June 1992 (23.06.92) US 080,386 21 June 1993 (21.06.93) US (71) Applicant: THE MT. SINAI SCHOOL OF MEDICINE OF THE CITY UNIVERSITY OF NEW YORK [US/ US]; One Gustave Levy Place, New York, NY 10029 (US). (72) Inventor: SEALFON, Stuart, C. ; 34 Sidney Place, Brook- lyn Heights, NY 11201 (US).		(74) Agents: CORUZZI, Laura, A. et al.; Pennie & Edmonds, 1155 Avenue of the Americas, New York, NY 10036 (US). (81) Designated States: AU, BB, BG, BR, BY, CA, CZ, FI, HU, JP, KR, KZ, LK, MG, MN, MW, NO, NZ, PL, RO, RU, SD, SK, UA, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i>
(54) Title: CLONING AND EXPRESSION OF GONADOTROPIN-RELEASING HORMONE RECEPTOR (57) Abstract The present invention relates to the GnRH-R genes and proteins. The DNA sequences disclosed herein may be engineered into expression systems designed for the production of GnRH-R and/or cell lines which express the GnRH-R and preferably respond to GnRH induced signal transduction. Such cell lines may advantageously be used for screening and identifying GnRH agonists and antagonists. In accordance with another aspect of the invention, the GnRH DNA, antisense oligonucleotide sequences, the GnRH expression products, and antibodies to such products may be used in the diagnosis and therapy of reproductive disorders associated with abnormal expression of the GnRH-R; e.g., overexpression, underexpression or expression or expression of a dysfunctional mutant receptor. Transgenic animals containing the GnRHR transgene may be used as animal models for the evaluation of GnRH analogs <i>in vivo</i> .		

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CLONING AND EXPRESSION OF
GONADOTROPIN-RELEASING HORMONE RECEPTOR

1. INTRODUCTION

5 The present invention relates to the cloning of
gonadotropin-releasing hormone receptor (GnRH-R), and
genetically engineered host cells which express the
GnRH-R. Such engineered cells may be used to evaluate
and screen drugs and analogs of GnRH involved in GnRH-
10 R activation, regulation and uncoupling.

2. BACKGROUND OF THE INVENTION

 The GnRH-R is a key mediator in the integration
of the neural and endocrine systems. Normal
15 reproduction depends on the pulsatile release of
physiological concentrations of GnRH which binds to
specific high affinity pituitary receptors and
triggers the secretion of the gonadotropins
luteinizing hormone (LH) and follicle stimulating
20 hormone (FSH). Whereas physiological concentrations
of GnRH orchestrate normal reproduction, high levels
of agonist lead to an opposite response, the
suppression of gonadotropin secretion. The capacity
of GnRH analogues both to activate and to inhibit the
25 hypothalamic-pituitary-gonadal axis has led to their
wide clinical utility in the treatment of a variety of
disorders ranging from infertility to prostatic
carcinoma.

 The responsiveness and capacity of the
30 gonadotrope GnRH-R is influenced by agonist,
concentration and pattern of exposure (Clayton, 1989,
J Endocrinol 120: 11-19). Both in vivo and in vitro
studies have demonstrated that low concentration
pulsatile GnRH is trophic to the receptor and that a
35 high concentration of agonist induces receptor down-
regulation and desensitization. The binding of GnRH

to its receptor stimulates phospholipase C and generates inositol-1,4,5-triphosphate and diacylglycerol (Huckle & Conn, 1988, Endocrine Reviews 9: 387-395). These second messengers, in turn, release calcium from intracellular stores and activate protein kinase C. Receptor up-regulation appears to involve both protein kinase C and calcium (Huckle & Conn, 1988, Endocrine Reviews 9: 387-395; Huckle et al., 1988, Journal of Biological Chemistry 263: 3296-3302; Young et al., 1985, Journal of Endocrinology 107: 49-56). It is not certain which effectors underlie down-regulation.

While great progress has been made in understanding the mechanisms underlying GnRH-R regulation and desensitization through receptor binding studies, direct measurement of GnRH-R gene transcription and biosynthesis has not been possible. Cloning of the GnRH-R cDNA would advance the evaluation of GnRH-R activation, regulation and uncoupling. Determining the primary sequence of the receptor would facilitate the directed design of improved analogues. However, despite intensive interest, heretofore, the GnRH-R gene has not been cloned and expressed in any species.

25

3. SUMMARY OF THE INVENTION

The present invention relates to the GnRH-R genes and proteins. The DNA sequences disclosed herein may be engineered into expression systems designed for the production of GnRH-R and/or cell lines which express the GnRH-R and preferably respond to GnRH induced signal transduction. Such cell lines may advantageously be used for screening and identifying GnRH agonists and antagonists. In accordance with another aspect of the invention, the GnRH DNA,

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antisense oligonucleotide sequences, the GnRH expression products, and antibodies to such products may be used in the diagnosis and therapy of reproductive disorders associated with abnormal expression of the GnRH-R; e.g., overexpression, underexpression or expression of a dysfunctional mutant receptor. Transgenic animals containing the GnRH-R transgene may be used as animal models for the evaluation of GnRH analogs in vivo.

10 The elucidation of the GnRH-R sequence described herein reflects a major advance in reproductive endocrinology and reveals the complex nature of GnRH-R signal transduction and regulation. Unlike most hormonal signals, GnRH is released in a pulsatile

15 fashion, with the frequency and amplitude of the pulses conveying crucial information (Weiss et al., 1990, Mol. Endocrinol. 4: 557-564; Hasenleder et al., 1991, Endocrinology 128: 509-517). GnRH-R binding capacity itself is either up- or down-regulated by

20 agonists depending on duration of exposure and concentration (Loumaye & Catt, 1982, Science 215: 983-985). The clinical utility of GnRH agonists, which help control a variety of human diseases, including prostatic hypertrophy, prostatic cancer, endometriosis

25 and precocious puberty, depends on this induction of pituitary desensitization. The cloning of the GnRH-R will lead to greater understanding of the complex interplay of hypothalamic, pituitary and gonadal hormones which underlies both pharmacotherapy and

30 reproduction.

4. DESCRIPTION OF THE FIGURES

Figure 1. Hybrid-arrest of serotonin (5HT) receptor and GnRH-R expression by antisense

35 oligonucleotides. 100 nM 5HT or 200 nM GnRH were

introduced into the bath at the horizontal lines. A, Response to 5HT and GnRH in oocytes previously injected with a mixture of rat brain RNA (for the 5HT response), α T3-1 RNA (for the GnRH response) and antisense 5HT_{1C} receptor oligonucleotide. 16 cells showed identical responses. B, Response to GnRH and 5HT in oocytes previously injected with a mixture of rat brain RNA, α T3-1 RNA and antisense WZ7 oligonucleotide. 24 cells had identical responses.

Figure 2. Characterization of clone WZ25 expressed in oocytes. A, Electrophysiological response to GnRH of oocytes injected with the WZ25 transcript in the absence (left) or presence (right) of GnRH antagonist. The three tracings shown are from different cells. Solid and dotted lines indicate GnRH and GnRH antagonist administration, respectively. Uninjected oocytes had no response to GnRH (n=12). B, Displacement of ¹²⁵I-GnRH-A by GnRH-A and GnRH in membranes of oocytes injected with transcript from WZ25. A comparative displacement curve using α T3-1 cell membranes combined with membranes from uninjected oocytes is also shown (O). Error bars show SEM.

Figure 3. Nucleotide (SEQ. ID NO: 1) and deduced amino acid sequences (SEQ. ID NO: 2) of clone WZ25. Numbering begins with the first methionine of the 981 base pair open reading frame. The deduced amino acid sequence is shown below the nucleotide sequence. Putative transmembrane regions I-VII are underlined. Symbols below the amino acid sequences indicate potential N-glycosylation sites (Δ), and phosphorylation sites for protein kinase A (\diamond), Casein kinase 2 (O) and protein kinase C (*) (Hubbard & Ivatt, 1981, Ann Rev. Biochem. 50: 555-583; Kemp & Pearson, 1990, Trends Biochem. Sci. 15: 342-346; Pearson & Kemp, 1991, Meth. Enzymol. 200: 62-81;

Kennelly & Krebs, 1991, J. Biol. Chem. 266: 15555-15558).

Figure 4. Hydrophobicity plot of the GnRH-R and amino acid sequence alignment of: GnR, mouse
5 gonadotropin-releasing hormone receptor; ILR, human interleukin-8 receptor (Murphy & Tiffany, 1991, Science 253: 1280-1283); SPR, rat substance P receptor (Hershey & Krause, 1990, Science 247: 958-962); β 1R, human β 1-adrenergic (Frielle et al., 1987, Proc. Natl.
10 Acad. Sci. USA 84: 7920-7924); and RHO, human rhodopsin (Nathans & Hogness, 1984, Proc. Natl. Acad. Sci. USA 81: 4851-4855). I-VII denote putative transmembrane regions. Boxes indicate identical amino acid residues.

15 Figure 5. Distribution of GnRH-R mRNA. Autoradiogram of A, solution hybridization assay using 2 μ g of total mouse pituitary, GT-1, GH3, and AtT20 RNA and 625 ng of α T3-1 total RNA, B, northern blot analysis with 3 μ g of poly(A)⁺ α T3-1 RNA, and C-F, rat
20 anterior pituitary in situ hybridization. C, antisense probe X-ray film autoradiography. D, sense probe control (calibration bar = 450 μ m). E,F, dark-field (calibration bar = 50 μ m), bright-field (calibration bar = 100 μ m) photomicrographs of
25 emulsion-dipped anterior pituitary section. The molecular weight markers are Hind III digested λ DNA.

Fig. 6. Expression of the human GnRH-R cDNA in *Xenopus* oocytes.

Fig. 7. Displacement of [¹²⁵I]GnRh agonist binding
30 to membranes prepared from COS-1 cells transfected with the pSV2A-human Gn-RHR construct.

Fig. 8. Effects of GnRH and GnRH antagonist on inositol phosphate production in COS-1 cells transfected with pSV2A-human GnRH-R.

Fig. 9. Nucleotide and putative amino acid sequence of the human GnRH-R.

Fig. 10. Northern blot analysis with human GnRH-R cDNA: lane 1(T): human testis poly(A) RNA; lane 2
5 (P): human pituitary poly(A) RNA; lane 3 (A): human β -actin cDNA; lane 4(R): human GnRH-R cDNA.

Fig. 11. Schematic of human GnRH-R.

5. DETAILED DESCRIPTION OF THE INVENTION

10

The present invention relates to the cloning and expression of murine and human GnRH-R. The GnRH-R, which plays a pivotal role in the reproductive system, is characterized by seven transmembrane domains
15 characteristic of G protein-coupled receptors, but lacks a typical intracellular C-terminus. The unusual structure and regulatory domain of the GnRH-R is responsible for the unique aspects of signal transduction and regulation mediated by the receptor.
20 The GnRH-R produced herein may be used to evaluate and screen drugs and analogs of GnRH involved in receptor activation, regulation and uncoupling. Alternatively, GnRH-R DNA, oligonucleotides and/or antisense sequences, or the GnRH-R, peptide fragments thereof,
25 or antibodies thereto may be used in the diagnosis and/or treatment of reproductive disorders.

For clarity of discussion, the invention is described in the subsections below by way of example for the murine and human GnRH-R. However, the
30 principles may be analogously applied to clone and express the GnRH-R of other species, and to clone and express other receptors belonging to the unique GnRH family, *i.e.*, G-protein type of receptors which lack an intracellular C-terminus and bind to GnRH or
35 analogs thereof.

5.1. THE GnRH-R CODING SEQUENCE

The nucleotide coding sequence (SEQ. ID NO: 1) and deduced amino acid sequence (SEQ. ID NO: 2) for the murine GnRH-R are depicted in Figure 3. The
5 longest open reading frame encodes a 327 amino acid protein of about 37,000 MW. Three consensus N-linked glycosylation sites are present, two in the N-terminus and one in the first extracellular loop (FIG. 3). Hydrophobicity analysis of the deduced protein reveals
10 seven stretches of highly hydrophobic amino acids with 20-30% sequence similarity to other G-protein receptors, with the highest degree of homology to the interleukin-8 receptor (FIG. 4).

The GnRH-R is nearly the smallest member of the
15 G-protein receptor superfamily, the first cytoplasmic loop of the GnRH-R is longer than any other G-protein receptor, and unlike any other G-protein receptor, it lacks a polar cytoplasmic C-terminus. While highly conserved residues are present in the GnRH-R, such as
20 the cysteines in each of the first two extracellular loops which stabilize many receptors, several features of the GnRH-R are unusual. For example, the highly conserved transmembrane II aspartate/glutamate, which has been found to be essential for the function of
25 many G-protein receptors, is replaced by asparagine. Another deviation from other G-protein receptors is the substitution of a serine for the conserved tyrosine located adjacent to transmembrane III. This creates a potential phosphorylation site, unique to
30 the GnRH-R, in a domain critical for signal transduction of other G-protein receptors. Other potential regulatory phosphorylation sites are also present (see FIG. 3).

The invention also relates to GnRH-R genes
35 isolated from other species, including humans. The

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human GnRH receptor was cloned by probing a λ gt10 human pituitary cDNA library with the mouse GnRH receptor insert which had been ^{32}P -labeled via random hexamer priming. To confirm that the isolated clone
5 encoded a functional human GnRH-R, synthetic RNA transcripts were injected into oocytes. All RNA-injected oocytes developed large depolarizing currents upon exposure to GnRH indicating that the cloned DNA fragment encoded a functional receptor.

10 The nucleotide coding sequence (SEQ. ID. NO:3) and deduced amino acid sequence (SEQ. ID. NO:4) for the human GnRH-R are depicted in Figure 9. Sequencing of the human clone identified a 2160 bp insert containing a 984 bp open reading frame. The open
15 reading frame encodes a 328 amino acid protein with 90% identity to the predicted sequence of the mouse receptor.

Hydrophobicity analysis identified the seven hydrophobic domains characteristics of G-protein
20 coupled receptors. As was found for the predicted structure of the mouse receptor, the human GnRH-R lacks essentially any C-terminal intracellular domain. Two potential N-linked glycosylation sites are present, one in each of the first extracellular
25 domains. Several cytoplasmic serine and threonine residues are found on intracellular domains and may serve as regulatory phosphorylation sites (FIG. 11).

Northern blot analysis, utilizing radioactively labelled human GnRH-R as a probe, identified a
30 transcript of roughly 4.7 kb in human pituitary poly(A) RNA (FIG. 10). No signal was detected in poly(A) RNA purified from human testis or with a human β -actin cDNA control.

To determine the extent of the 5' and 3'-
35 untranslated domains of the RNA, PCR analysis of the

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phage isolates from the primary library screening was undertaken. An antisense oligonucleotide primer representing sequence near the 5'-end of the GnRH-R cDNA insert or a sense primer near the 3'-end of the same sequence was used in conjunction with primers designed against the adjacent GTI-cloning site to map the unpurified clones. The longest PCR products identified had -1.3 kb of additional 5'-sequence and -0.3 kb of additional 3'-sequence. These data suggest that the GnRH-R mRNA contains at least 1.3 kb of 5'-untranslated sequence and 1.5 kb of 3'-untranslated sequence. Based on the Northern blot data, this suggests that additional untranslated sequence (<1 kb) is not contained in any of the clones isolated.

The invention also relates to GnRH-R genes isolated from other species in which GnRH-R activity exists. Members of the GnRH-R family are defined herein as those receptors that bind GnRH. Such receptors may demonstrate about 80% homology at the nucleotide level, and even 90% homology at the amino acid level in substantial stretches of sequences located in regions outside the transmembrane domains.

Cloning of other receptors in the GnRH-R family may be carried out in a number of different ways. For example, the murine and human sequence can be used to design degenerate or fully degenerate oligonucleotide probes which can be used as PCR probes or to screen cDNA libraries derived from appropriate cells which express the GnRH-R, or genomic libraries. The N-terminus and cytoplasmic loops (both intracellular and extracellular) of the murine and human sequences depicted in FIG. 3 and FIG. 11, respectively, may advantageously be used to design such oligonucleotide probes, as these regions should be relatively conserved within the GnRH-R family.

Alternatively, a bacteriophage cDNA library may be screened, under conditions of reduced stringency, using a radioactively labeled fragment of the human or murine GnRH-R clone to isolate GnRH-R related
5 proteins. For a review of cloning strategies which may be used, see E.g., Maniatis, 1989, Molecular Cloning, A Laboratory Manual, Cold Springs Harbor Press, N.Y.; and Ausubel et al., 1989, Current
10 Protocols in Molecular Biology, Green Publishing Associates and Wiley Interscience, N.Y.

In accordance with the invention, nucleotide sequences which encode a GnRH-R, fragments, fusion proteins or functional equivalents thereof, may be used to generate recombinant DNA molecules that direct
15 the expression of the GnRH-R, or a functionally active peptide, fusion protein or functional equivalent thereof, in appropriate host cells. Alternatively, nucleotide sequences which hybridize to portions of the GnRH-R sequence may also be used in nucleic acid
20 hybridization assays, Southern and Northern blot analyses, etc.

Due to the degeneracy of the genetic code, other DNA sequences which encode substantially the GnRH-R amino acid sequence, e.g., such as the murine sequence
25 (SEQ. ID NO: 2) depicted in FIG. 3 or the human sequence, or a functional equivalent may be used in the practice of the present invention for the cloning and expression of the GnRH-R. Such DNA sequences include those which are capable of hybridizing to the
30 murine or human GnRH-R sequence under stringent conditions, or which would be capable of hybridizing under stringent conditions but for the degeneracy of the genetic code. The stringency conditions may be adjusted in a number of ways. For example, when
35 performing polymerase chain reactions (PCR), the

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- temperature at which annealing of primers to template takes place or the concentration of $MgCl_2$ in the reaction buffer may be adjusted. When using radioactively labeled DNA fragments or
- 5 oligonucleotides to probe filters, the stringency may be adjusted by changes in the ionic strength of the wash solutions or by carefully controlling the temperature at which the filter washes are carried out.
- 10 Altered DNA sequences which may be used in accordance with the invention include deletions, additions or substitutions of different nucleotide residues resulting in a sequence that encodes the same or a functionally equivalent gene product. The gene
- 15 product itself may contain deletions, additions or substitutions of amino acid residues within the GnRH-R sequence, which result in a silent change thus producing a functionally equivalent GnRH-R. Such amino acid substitutions may be made on the basis of
- 20 similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipatic nature of the residues involved. For example, negatively charged amino acids include aspartic acid and glutamic acid; positively charged amino acids
- 25 include lysine and arginine; amino acids with uncharged polar head groups having similar hydrophilicity values include the following: leucine, isoleucine, valine; glycine, aniline; asparagine, glutamine; serine, threonine; phenylalanine, tyrosine.
- 30 As used herein, a functionally equivalent GnRH-R refers to a receptor which binds to GnRH, but not necessarily with the same binding affinity of its counterpart native GnRH-R.

The DNA sequences of the invention may be

35 engineered in order to alter the GnRH-R coding

- sequence for a variety of ends including but not limited to alterations which modify processing and expression of the gene product. For example, mutations may be introduced using techniques which are
- 5 well known in the art, e.g. site-directed mutagenesis, to insert new restriction sites, to alter glycosylation patterns, phosphorylation, etc. For example, in certain expression systems such as yeast, host cells may over glycosylate the gene product.
- 10 When using such expression systems it may be preferable to alter the GnRH-R coding sequence to eliminate the N-linked glycosylation site; e.g. in the murine sequence this may be accomplished by altering one or more glycosylation sites indicated in FIG. 3.
- 15 In another embodiment for the invention, the GnRH-R or a modified GnRH-R sequence may be ligated to a heterologous sequence to encode a fusion protein. The fusion protein may be engineered to contain a cleavage site located between the GnRH-R sequence and the
- 20 heterologous protein sequence, so that the GnRH-R can be cleaved away from the heterologous moiety.
- In an alternate embodiment of the invention, the coding sequence of GnRH-R could be synthesized in whole or in part, using chemical methods well known in
- 25 the art. See, for example, Caruthers, et al., 1980, Nuc. Acids Res. Symp. Ser. 7:215-233; Crea and Horn, 180, Nuc. Acids Res. 9(10):2331; Matteucci and Caruthers, 1980, Tetrahedron Letters 21:719; and Chow and Kempe, 1981, Nuc. Acids Res. 9(12):2807-2817.
- 30 Alternatively, the protein itself could be produced using chemical methods to synthesize the GnRH-R amino acid sequence in whole or in part. For example, peptides can be synthesized by solid phase techniques, cleaved from the resin, and purified by preparative
- 35 high performance liquid chromatography. (E.g., see

Creighton, 1983, Proteins Structures And Molecular Principles, W.H. Freeman and Co., N.Y. pp. 50-60). The composition of the synthetic peptides may be confirmed by amino acid analysis or sequencing (e.g.,
5 the Edman degradation procedure; see Creighton, 1983, Proteins, Structures and Molecular Principles, W.H. Freeman and Co., N.Y., pp. 34-49).

5.2. EXPRESSION OF THE GnRH-R

10 In order to express a biologically active GnRH-R, the nucleotide sequence coding for GnRH-R, or a functional equivalent as described in Section 5.1 supra, is inserted into an appropriate expression vector, i.e., a vector which contains the necessary
15 elements for the transcription and translation of the inserted coding sequence. The GnRH-R gene products as well as host cells or cell lines transfected or transformed with recombinant GnRH-R expression vectors can be used for a variety of purposes. These include
20 but are not limited to generating antibodies (i.e., monoclonal or polyclonal) that bind to the receptor, including those that competitively inhibit GnRH binding and "neutralize" GnRH activity; the screening and selection of GnRH analogs or drugs that act via
25 the GnRH-R; etc.

5.2.1. EXPRESSION SYSTEMS

Methods which are well known to those skilled in the art can be used to construct expression vectors
30 containing the GnRH-R coding sequence and appropriate transcriptional/translational control signals. These methods include in vitro recombinant DNA techniques, synthetic techniques and in vivo recombination/genetic recombination. See, for example, the techniques
35 described in Maniatis et al., 1989, Molecular Cloning

A Laboratory Manual, Cold Spring Harbor Laboratory, N.Y. and Ausubel et al., 1989, Current Protocols in Molecular Biology, Greene Publishing Associates and Wiley Interscience, N.Y.

- 5 A variety of host-expression vector systems may be utilized to express the GnRH-R coding sequence. These include but are not limited to microorganisms such as bacteria transformed with recombinant bacteriophage DNA, plasmid DNA or cosmid DNA
- 10 expression vectors containing the GnRH-R coding sequence; yeast transformed with recombinant yeast expression vectors containing the GnRH-R coding sequence; insect cell systems infected with recombinant virus expression vectors (e.g.,
- 15 baculovirus) containing the GnRH-R coding sequence; plant cell systems infected with recombinant virus expression vectors (e.g., cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or transformed with recombinant plasmid expression vectors (e.g., Ti
- 20 plasmid) containing the GnRH-R coding sequence; or animal cell systems infected with recombinant virus expression vectors (e.g., adenovirus, vaccinia virus) including cell lines engineered to contain multiple copies of the GnRH-R DNA either stably amplified
- 25 (e.g., CHO/dhfr) or unstably amplified in double-minute chromosomes (e.g., murine cell lines).

The expression elements of these systems vary in their strength and specificities. Depending on the host/vector system utilized, any of a number of

30 suitable transcription and translation elements, including constitutive and inducible promoters, may be used in the expression vector. For example, when cloning in bacterial systems, inducible promoters such as pL of bacteriophage λ , plac, ptrp, ptac (ptrp-lac

35 hybrid promoter) and the like may be used; when

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cloning in insect cell systems, promoters such as the baculovirus polyhedrin promoter may be used; when cloning in plant cell systems, promoters derived from the genome of plant cells (e.g., heat shock promoters; 5 the promoter for the small subunit of RUBISCO; the promoter for the chlorophyll a/b binding protein) or from plant viruses (e.g., the 35S RNA promoter of CaMV; the coat protein promoter of TMV) may be used; when cloning in mammalian cell systems, promoters 10 derived from the genome of mammalian cells (e.g., metallothionein promoter) or from mammalian viruses (e.g., the adenovirus late promoter; the vaccinia virus 7.5K promoter) may be used; when generating cell lines that contain multiple copies of the GnRH-R DNA 15 SV40-, BPV- and EBV-based vectors may be used with an appropriate selectable marker.

In bacterial systems a number of expression vectors may be advantageously selected depending upon the use intended for the GnRH-R expressed. For 20 example, when large quantities of GnRH-R are to be produced for the generation of antibodies, vectors which direct the expression of high levels of fusion protein products that are readily purified may be desirable. Such vectors include but are not limited 25 to the E. coli expression vector pUR278 (Ruther et al., 1983, EMBO J. 2:1791), in which the GnRH-R coding sequence may be ligated into the vector in frame with the lac Z coding region so that a hybrid AS-lac Z protein is produced; pIN vectors (Inouye & Inouye, 30 1985, Nucleic acids Res. 13:3101-3109; Van Heeke & Schuster, 1989, J. Biol. Chem. 264:5503-5509); and the like. pGEX vectors may also be used to express foreign polypeptides as fusion proteins with 35 glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified

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from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. The pGEX vectors are designed to include thrombin or factor Xa protease cleavage sites so that
5 the cloned polypeptide of interest can be released from the GST moiety.

In yeast, a number of vectors containing constitutive or inducible promoters may be used. For a review see, Current Protocols in Molecular Biology,
10 Vol. 2, 1988, Ed. Ausubel et al., Greene Publish. Assoc. & Wiley Interscience, Ch. 13; Grant et al., 1987, Expression and Secretion Vectors for Yeast, in Methods in Enzymology, Eds. Wu & Grossman, 1987, Acad. Press, N.Y., Vol. 153, pp. 516-544; Glover, 1986, DNA
15 Cloning, Vol. II, IRL Press, Wash., D.C., Ch. 3; and Bitter, 1987, Heterologous Gene Expression in Yeast, Methods in Enzymology, Eds. Berger & Kimmel, Acad. Press, N.Y., Vol. 152, pp. 673-684; and The Molecular Biology of the Yeast *Saccharomyces*, 1982, Eds.
20 Strathern et al., Cold Spring Harbor Press, Vols. I and II.

In cases where plant expression vectors are used, the expression of the GnRH-R coding sequence may be driven by any of a number of promoters. For example,
25 viral promoters such as the 35S RNA and 19S RNA promoters of CaMV (Brisson et al., 1984, Nature 310:511-514), or the coat protein promoter of TMV (Takamatsu et al., 1987, EMBO J. 6:307-311) may be used; alternatively, plant promoters such as the small
30 subunit of RUBISCO (Coruzzi et al., 1984, EMBO J. 3:1671-1680; Broglie et al., 1984, Science 224:838-843); or heat shock promoters, e.g., soybean hsp17.5-E or hsp17.3-B (Gurley et al., 1986, Mol. Cell. Biol. 6:559-565) may be used. These constructs can be
35 introduced into plant cells using Ti plasmids, Ri

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plasmids, plant virus vectors, direct DNA transformation, microinjection, electroporation, etc. For reviews of such techniques see, for example, Weissbach & Weissbach, 1988, Methods for Plant

- 5 Molecular Biology, Academic Press, NY, Section VIII, pp. 421-463; and Grierson & Corey, 1988, Plant Molecular Biology, 2d Ed., Blackie, London, Ch. 7-9.

- An alternative expression system which could be used to express GnRH-R is an insect system. In one
10 such system, Autographa californica nuclear polyhidrosis virus (AcNPV) is used as a vector to express foreign genes. The virus grows in Spodoptera frugiperda cells. The GnRH-R coding sequence may be cloned into non-essential regions (for example the
15 polyhedrin gene) of the virus and placed under control of an AcNPV promoter (for example the polyhedrin promoter). Successful insertion of the GnRH-R coding sequence will result in inactivation of the polyhedrin gene and production of non-occluded recombinant virus
20 (i.e., virus lacking the proteinaceous coat coded for by the polyhedrin gene). These recombinant viruses are then used to infect Spodoptera frugiperda cells in which the inserted gene is expressed. (E.g., see Smith et al., 1983, J. Virol. 46:584; Smith, U.S.
25 Patent No. 4,215,051).

- In mammalian host cells, a number of viral based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, the GnRH-R coding sequence may be ligated to an adenovirus
30 transcription/translation control complex, e.g., the late promoter and tripartite leader sequence. This chimeric gene may then be inserted in the adenovirus genome by in vitro or in vivo recombination. Insertion in a non-essential region of the viral
35 genome (e.g., region E1 or E3) will result in a

recombinant virus that is viable and capable of expressing GnRH-R in infected hosts. (E.g., See Logan & Shenk, 1984, Proc. Natl. Acad. Sci. (USA) 81:3655-3659). Alternatively, the vaccinia 7.5K promoter may
5 be used. (E.g., see Mackett et al., 1982, Proc. Natl. Acad. Sci. (USA) 79:7415-7419; Mackett et al., 1984, J. Virol. 49:857-864; Panicali et al., 1982, Proc. Natl. Acad. Sci. 79:4927-4931).

Specific initiation signals may also be required
10 for efficient translation of inserted GnRH-R coding sequences. These signals include the ATG initiation codon and adjacent sequences. In cases where the entire GnRH-R gene, including its own initiation codon and adjacent sequences, is inserted into the
15 appropriate expression vector, no additional translational control signals may be needed. However, in cases where only a portion of the GnRH-R coding sequence is inserted, exogenous translational control signals, including the ATG initiation codon, must be
20 provided. Furthermore, the initiation codon must be in phase with the reading frame of the GnRH-R coding sequence to ensure translation of the entire insert. These exogenous translational control signals and initiation codons can be of a variety of origins, both
25 natural and synthetic. The efficiency of expression may be enhanced by the inclusion of appropriate transcription enhancer elements, transcription terminators, etc. (see Bitter et al., 1987, Methods in Enzymol. 153:516-544).

30 In addition, a host cell strain may be chosen which modulates the expression of the inserted sequences, or modifies and processes the gene product in the specific fashion desired. Such modifications (e.g., glycosylation) and processing (e.g., cleavage)
35 of protein products may be important for the function

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of the protein. Different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins. Appropriate cells lines or host systems can be chosen
5 to ensure the correct modification and processing of the foreign protein expressed. To this end, eukaryotic host cells which possess the cellular machinery for proper processing of the primary transcript, glycosylation, and phosphorylation of the
10 gene product may be used. Such mammalian host cells include but are not limited to CHO, VERO, BHK, HeLa, COS, MDCK, 293, WI38, etc.

For long-term, high-yield production of recombinant proteins, stable expression is preferred.
15 For example, cell lines which stably express the GnRH-R may be engineered. Rather than using expression vectors which contain viral origins of replication, host cells can be transformed with the GnRH-R DNA controlled by appropriate expression control elements
20 (e.g., promoter, enhancer, sequences, transcription terminators, polyadenylation sites, etc.), and a selectable marker. Following the introduction of foreign DNA, engineered cells may be allowed to grow for 1-2 days in an enriched media, and then are
25 switched to a selective media. The selectable marker in the recombinant plasmid confers resistance to the selection and allows cells to stably integrate the plasmid into their chromosomes and grow to form foci which in turn can be cloned and expanded into cell
30 lines. This method may advantageously be used to engineer cell lines which express the GnRH-R on the cell surface, and which respond to GnRH mediated signal transduction. Such engineered cell lines are particularly useful in screening GnRH analogs.

35

- 20 -

- A number of selection systems may be used, including but not limited to the herpes simplex virus thymidine kinase (Wigler, et al., 1977, Cell 11:223), hypoxanthine-guanine phosphoribosyltransferase (Szybalska & Szybalski, 1962, Proc. Natl. Acad. Sci. USA 48:2026), and adenine phosphoribosyltransferase (Lowy, et al., 1980, Cell 22:817) genes can be employed in tk⁻, hgp^rt⁻ or ap^rt⁻ cells, respectively. Also, antimetabolite resistance can be used as the basis of selection for dhfr, which confers resistance to methotrexate (Wigler, et al., 1980, Natl. Acad. Sci. USA 77:3567; O'Hare, et al., 1981, Proc. Natl. Acad. Sci. USA 78:1527); gpt, which confers resistance to mycophenolic acid (Mulligan & Berg, 1981, Proc. Natl. Acad. Sci. USA 78:2072); neo, which confers resistance to the aminoglycoside G-418 (Colberre-Garapin, et al., 1981, J. Mol. Biol. 150:1); and hyg^r, which confers resistance to hygromycin (Santerre, et al., 1984, Gene 30:147) genes.
- Recently, additional selectable genes have been described, namely trpB, which allows cells to utilize indole in place of tryptophan; hisD, which allows cells to utilize histinol in place of histidine (Hartman & Mulligan, 1988, Proc. Natl. Acad. Sci. USA 85:8047); and ODC (ornithine decarboxylase) which confers resistance to the ornithine decarboxylase inhibitor, 2-(difluoromethyl)-DL-ornithine, DFMO (McConlogue L., 1987, In: Current Communications in Molecular Biology, Cold Spring Harbor Laboratory ed.).
- In a specific embodiment, described herein, the human GnRH-R cDNA was subcloned into an expression vector, pSV2A, containing the SV40 early promoter. COS-1 cells were transiently transfected with the pSV2A-human GnRH-R construct using the DEAE-dextran method of transfection (Keown, W.A. et al., 1990, in

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Methods of Enzymology, Vl. 185 (Goeddel, D.V., ed.)
pg. 527-537 Academic Press, New York). Experiments,
using membranes from COS-1 transfected cells,
indicated that the heterologously expressed receptor
5 was capable of binding GnRH. Ligand binding was also
found to be coupled to inositol phosphate metabolism
indicating further that the transfected COS-1 cells
expressed a functional human GnRH-R.

10 5.2.2. IDENTIFICATION OF
TRANSFECTANTS OR TRANSFORMANTS
 THAT EXPRESS THE GnRH-R

The host cells which contain the coding sequence
and which express the biologically active gene product
may be identified by at least four general approaches;
15 (a) DNA-DNA or DNA-RNA hybridization; (b) the presence
or absence of "marker" gene functions; (c) assessing
the level of transcription as measured by the
expression of GnRH-R mRNA transcripts in the host
cell; and (d) detection of the gene product as
20 measured by immunoassay or by its biological activity.

In the first approach, the presence of the GnRH-R
coding sequence inserted in the expression vector can
be detected by DNA-DNA or DNA-RNA hybridization using
probes comprising nucleotide sequences that are
25 homologous to the GnRH-R coding sequence,
respectively, or portions or derivatives thereof.

In the second approach, the recombinant
expression vector/host system can be identified and
selected based upon the presence or absence of certain
30 "marker" gene functions (e.g., thymidine kinase
activity, resistance to antibiotics, resistance to
methotrexate, transformation phenotype, occlusion body
formation in baculovirus, etc.). For example, if the
GnRH-R coding sequence is inserted within a marker
35 gene sequence of the vector, recombinants containing

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the GnRH-R coding sequence can be identified by the absence of the marker gene function. Alternatively, a marker gene can be placed in tandem with the GnRH-R sequence under the control of the same or different
5 promoter used to control the expression of the GnRH-R coding sequence. Expression of the marker in response to induction or selection indicates expression of the GnRH-R coding sequence.

In the third approach, transcriptional activity
10 for the GnRH-R coding region can be assessed by hybridization assays. For example, RNA can be isolated and analyzed by Northern blot using a probe homologous to the GnRH-R coding sequence or particular portions thereof. Alternatively, total nucleic acids
15 of the host cell may be extracted and assayed for hybridization to such probes.

In the fourth approach, the expression of the GnRH-R protein product can be assessed immunologically, for example by Western blots,
20 immunoassays such as radioimmuno-precipitation, enzyme-linked immunoassays and the like. The ultimate test of the success of the expression system, however, involves the detection of the biologically active GnRH-R gene product. A number of assays can be used
25 to detect receptor activity including but not limited to GnRH binding assays; and GnRH biological assays using engineered cell lines as the test substrate.

In a specific embodiment described herein, cell membranes were prepared from COS-1 cells transfected
30 with a recombinant expression vector containing the human GnRH-R cDNA. Expression of the human GnRH-R was detected using a ¹²⁵I labeled GnRH analog. In addition the expression of biologically active GnRH-R could be detected in transfected cells by measuring levels of

35

GnRH-stimulated inositol phosphate (IP) production as described in Section 7.1.5.

5.2.3. RECOVERY OF THE GnRH-R

5 Once a clone that produces high levels of
biologically active GnRH-R is identified, the clone
may be expanded and used to produce large amounts of
the receptor which may be purified using techniques
well-known in the art including, but not limited to
10 immunoaffinity purification, chromatographic methods
including high performance liquid chromatography,
affinity chromatography using immobilized ligand such
as GnRH or analogs thereof bound to beads,
immunoaffinity purification using antibodies and the
15 like.

Where the GnRH-R coding sequence is engineered to
encode a cleavable fusion protein, purification may be
readily accomplished using affinity purification
techniques. For example, a collagenase cleavage
20 recognition consensus sequence may be engineered
between the carboxy terminus of GnRH-R and protein A.
The resulting fusion protein may be readily purified
using an IgG column that binds the protein A moiety.
Unfused GnRH-R may be readily released from the column
25 by treatment with collagenase. Another example would
be the use of pGEX vectors that express foreign
polypeptides as fusion proteins with glutathione S-
transferase (GST). The fusion protein may be
engineered with either thrombin or factor Xa cleavage
30 sites between the cloned gene and the GST moiety. The
fusion protein may be easily purified from cell
extracts by adsorption to glutathione agarose beads
followed by elution in the presence of glutathione.
In this aspect of the invention, any cleavage site or
35 enzyme cleavage substrate may be engineered between

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the GnRH-R sequence and a second peptide or protein that has a binding partner which could be used for purification, e.g., any antigen for which an immunoaffinity column can be prepared.

5

5.3. GENERATION OF ANTIBODIES THAT DEFINE THE GnRH-R

Various procedures known in the art may be used for the production of antibodies to epitopes of the recombinantly produced GnRH-R. Neutralizing
10 antibodies i.e., those which compete for the GnRH binding site of the receptor are especially preferred for diagnostics and therapeutics. Antibodies which define viral serological markers would be preferred
15 for diagnostic uses. Such antibodies include but are not limited to polyclonal, monoclonal, chimeric, single chain, Fab fragments and fragments produced by an Fab expression library.

For the production of antibodies, various host
20 animals may be immunized by injection with the GnRH-R including but not limited to rabbits, mice, rats, etc. Various adjuvants may be used to increase the immunological response, depending on the host species, including but not limited to Freund's (complete and
25 incomplete), mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, dinitrophenol, and potentially useful human adjuvants such as BCG
30 (bacilli Calmette-Guerin) and corynebacterium parvum.

Monoclonal antibodies to GnRH-R may be prepared by using any technique which provides for the production of antibody molecules by continuous cell
lines in culture. These include but are not limited
35 to the hybridoma technique originally described by Kohler and Milstein, (Nature, 1975, 256:495-497), the

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human B-cell hybridoma technique (Kosbor et al., 1983, Immunology Today, 4:72; Cote et al., 1983, Proc. Natl. Acad. Sci., 80:2026-2030) and the EBV-hybridoma technique (Cole et al., 1985, Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., pp. 77-96).
5 In addition, techniques developed for the production of "chimeric antibodies" (Morrison et al., 1984, Proc. Natl. Acad. Sci., 81:6851-6855; Neuberger et al., 1984, Nature, 312:604-608; Takeda et al., 1985,
10 Nature, 314:452-454) by splicing the genes from a mouse antibody molecule of appropriate antigen specificity together with genes from a human antibody molecule of appropriate biological activity can be used. Alternatively, techniques described for the
15 production of single chain antibodies (U.S. Patent 4,946,778) can be adapted to produce GnRH-R-specific single chain antibodies.

Antibody fragments which contain specific binding sites of GnRH-R may be generated by known techniques.
20 For example, such fragments include but are not limited to: the F(ab')₂ fragments which can be produced by pepsin digestion of the antibody molecule and the Fab fragments which can be generated by reducing the disulfide bridges of the F(ab')₂ fragments.
25 Alternatively, Fab expression libraries may be constructed (Huse et al., 1989, Science, 246:1275-1281) to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity to GnRH-R.

30

5.4. USES OF THE GnRH-R, DNA AND ENGINEERED CELL LINES

The GnRH-R DNA, antisense oligonucleotides, GnRH-R expression products, antibodies and engineered
35 cell lines described above have a number of uses for

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the diagnosis and treatment of reproductive disorders and in drug design and discovery.

For example, the GnRH-R DNA sequence may be used in hybridization assays of biopsies to diagnose
5 abnormalities of GnRH-R expression; e.g., Southern or Northern analysis, including in situ hybridization assays. In therapeutic applications, antisense or ribozyme molecules designed on the basis of the GnRH-R DNA sequence may be utilized to block transportation
10 and expression of the GnRH-R gene product. In this regard, oligonucleotides derived from the translation initiation site, e.g., between -10 and +10 regions of the GnRH-R nucleotide sequence, are preferred. Alternatively, the GnRH-R DNA could be used in gene
15 therapy approach to introduce the normal recombinant gene into the defective cells of an individual or to correct an endogenous mutation in order to reconstitute the GnRH-R and its function.

In another embodiment of the invention,
20 antibodies specific for the GnRH-R may be used to determine the pattern of receptor expression in biopsy tissue, or for diagnostic imaging in vivo; in such applications, "neutralizing" antibodies may be preferred. For example, an antibody conjugated to an
25 imaging compound could be administered to a patient to "map" the locations and distribution of the GnRH-R in vivo.

In another embodiment of the invention, the GnRH-R itself, or a fragment containing its GnRH
30 binding site, could be administered in vivo. The free GnRH-R or the peptide fragment could competitively bind to GnRH and inhibit its interaction with the native receptor in vivo.

In another embodiment of the invention,
35 stimulation of an antibody response, specific for

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GnRH-R, may be used as a means of contraception. For example, various host animals may be immunized by injection with GnRH-R or GnRH-R fusion protein, leading to stimulation of their immune system and
5 production of circulating anti-GnRH-R antibodies.

In yet another embodiment, the engineered cell lines which express the GnRH-R and respond to signal transduction may be utilized to screen and identify biologically active GnRH analogs, i.e., either
10 agonists or antagonists. Transgenic animals which contain the GnRH-R DNA as the transgene may be engineered to test the effects of such agonists or antagonists in vivo.

Recently, computer generated models for ligand-
15 receptor interactions have been developed and in a specific embodiment of the invention information derived from computer modeling of GnRH-R may be used for design of receptor agonist or antagonist. Over 74 distinct GPR (G-protein receptors) sequences have been
20 published and sequence alignments with GnRH-R sequences may facilitate understanding the role of certain protein sequences in determining ligand binding and regulation. Changes made to GnRH-R sequences, using for example techniques for site
25 directed mutagenesis, and expression of mutant receptors in cell lines may be used to further define the functional role of particular receptor regions and residues.

30 6. EXAMPLE: CLONING OF A FUNCTIONAL MURINE GnRH-R

The subsections below describe the cloning of a complementary DNA representing the mouse GnRH-R and confirm its identity using Xenopus oocyte expression.
35 Injection of sense RNA transcript leads to the expression of a functional, high-affinity GnRH-R.

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Expression of the GnRH-R using gonadotrope cell line RNA, however, is blocked by an antisense oligonucleotide. In situ hybridization in the rat anterior pituitary reveals a characteristic GnRH-R distribution. The nucleotide sequence encodes a 327 amino acid protein which has the seven putative transmembrane domains characteristic of G protein-coupled receptors, but which lacks a typical intracellular C-terminus. The unusual structure and novel potential regulatory domain of the GnRH-R may explain unique aspects of its signal transduction and regulation.

6.1. MATERIALS AND METHODS

Drugs were obtained from the following sources: the GnRH antagonist [D-Phe^{2,6},Pro³]-GnRH (Bachem, Torrance, CA), buserelin (D-Ser(But)⁶,Pro⁹-N-ethylamide GnRH) Hoechst-Roussel Pharmaceuticals (Somerville, NJ). All other chemicals were from Sigma Chemical Co. (St. Louis, MO). All animal care was in accordance with the NIH Guide for the Care and Use of Laboratory Animals.

6.1.1. OOCYTE MICRO-INJECTION AND RECORDING

Adult female *Xenopus laevis* (Nasco, Ft. Atkinson, WI) were kept at 18-20°C and a day/night cycle of 15h/9h. Oocytes were prepared for injection and the responses recorded as previously described (Sealfon et al., 1990, Mol. Endocrinol 4: 119-124). Cells were placed in a 0.5 ml bath and voltage clamped at -70 mV using standard two electrode technique (Dascal, 1987, CRC Crit. Rev. Biochem. 417: 47-61). Peptide ligands were diluted in the perfusion buffer and introduced into the bath. The clamp current was recorded using a chart recorder. Reversal potentials were determined by continuous ramping from -70 to +10 mV over 2

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seconds with and without agonist through an IBM PC/AT system using the TL-1 interface and pCLAMP software from Axon Instruments (Burlingame, CA).

5 6.1.2. PCR CLONING AND HYBRID
 ARREST SCREENING

RNA preparation and cDNA synthesis were performed as previously described (Sealfon et al., 1990, Mol. Endocrinol 4: 119-124; Snyder et al., 1991, Neurosci Lett 122: 37-40). Subclones for hybrid arrest
10 screening were isolated using PCR with a variety of degenerate oligonucleotides corresponding to conserved transmembrane domains of the GPR superfamily. The oligonucleotides used to isolate the group of
15 subclones including WZ7, modified from sequences of published oligomers (Zhou et al., 1990, Nature 347: 76-80), corresponded to transmembrane III (5'-GAGTCGACCTGTG(CT)G(CT)(GC)AT(CT)(AG)CNNT(GT)GAC(AC)G(CG)TAC-3') and transmembrane VI (5'-
20 CAGAATTCAG(AT)AGGGCANCCAGCAGAN(CG)(AG)(CT)GAA-3'). PCR was performed at low stringency. A portion of the reaction was reamplified at high stringency, digested with restriction enzymes, subcloned into pBluescript II KS+ (Stratagene) and sequenced. For hybrid-arrest
25 assay, an antisense oligonucleotide corresponding to transmembrane II of the 5HT_{1C} receptor (5'-ATCAGCAATGGCTAG-3') (Julius et al., 1988, Science 241: 558-564) and an oligonucleotide corresponding to WZ7 (5'-AGCATGATGAGGAGG-3') were synthesized. A
30 mixture of α T3-1 (1mg/ml) and rat brain total RNA (1mg/ml) was preincubated with antisense oligonucleotide (100 μ g/ml) for 10 minutes at 37°C in a buffer containing 200mM NaCl and 5mM Tris, pH 7.4 in a 3 μ l volume. Xenopus oocytes were injected with 50nl
35 of the mixture and incubated for 48 hours before recording.

- 30 -

6.1.3. LIBRARY SCREENING AND SEQUENCING

10⁶ plaques of a UniZap (Stratagene) αT3-1 cDNA library were screened with the insert of WZ7 which had been ³²P-labelled by random hexamer primers. 40

5 positive plaques were identified and 7 purified on secondary and tertiary screening. WZ25 was subcloned into pBluescript II SK+ by helper phage excision and both strands sequenced by the dideoxy-chain termination method with Sequenase T7 DNA polymerase

10 (USB). Sequence was further confirmed by resequencing both strands using taq polymerase labelling and an Applied Biosystems automated sequencer. To exclude the possibility that the predicted cytoplasmic C-

15 3' terminus was truncated due to a mutation in WZ25, the 3' sequence was confirmed in two additional independent clones. The nucleotide and amino acid sequence were analyzed using the Wisconsin GCG package on a VAX computer and MacVector (IBI) on a microprocessor.

20

6.1.4. CHARACTERIZATION OF WZ25RNA TRANSCRIPT

WZ25 in pBluescript II SK+ (Stratagene) was linearized and capped RNA transcript synthesized using

25 T3 RNA polymerase (Stratagene). Oocytes were injected with 1.25ng of the resulting transcript and incubated for 48 hours before recording. Oocytes were pre-treated with either buffer or a GnRH antagonist (antagonist 6: [Ac-D-Nal(2)¹,D,α-Me-pCl-Phe²,D-Trp³,D-Arg⁶,D-Ala¹⁰]GnRH; antagonist 27: [Ac-D-Nal(2)¹,D-α-Me-pCl-Phe²,D-Trp³,N-ε-lpr-Lys⁵,D-Tyr⁶,D-Ala¹⁰]GnRH; ref.

30 (Can der Spuy et al., 1987, In: Vickery BH and Nestor JJ (eds) LHRH and its Analogs: Contraceptive and Therapeutic Applications. NTP Press, Lancaster, England) for 3 minutes prior to GnRH administration.

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To confirm receptor expression, oocytes were re-exposed to GnRH after a three minute washout of antagonist.

5 6.1.5. RADIOLIGAND BINDING ASSAY

For membrane preparation, 500 oocytes were each injected with 2.5 ng synthetic WZ25 RNA. After 48 hours, oocyte membranes were prepared as described (Kobilka et al., 1987, J. Biol. Chem. 262: 15796-
10 15802) and resuspended in binding buffer containing 10mM HEPES, 1mM EDTA, and 0.1% bovine serum albumin to give a final concentration of 20 oocytes/ml. The receptor binding assay using ¹²⁵I-[D-Ala⁶, NaMe-Leu⁷, Pro⁹-NHET]GnRH (GnRH-A) was based on that previously
15 described for rat and sheep pituitary membranes (Millar et al., 1989, J. Biol. Chem. 264: 21007-21013). The binding in the presence of 10⁻⁶M GnRH analogue was considered to represent non-specific binding. Average B₀ (maximal binding) and non-
20 specific binding values were 1429 and 662 cpm, respectively. The dissociation constant (K_d) for GnRH-A and GnRH was determined using Enzfitter (Elsevier-BIOSOFT).

25 6.1.6. SOLUTION HYBRIDIZATION,
NORTHERN BLOT ANALYSIS,
AND IN SITU HYBRIDIZATION

A 399 nucleotide ³²P-labelled GnRH-R and a 117 nucleotide 1B15 (cyclophilin internal standard) antisense cRNA probe were synthesized and hybridized
30 to RNA in solution using described methods (Autelitano et al., 1989, Mol. Cell. Endo. 67: 101-105). Northern blot analysis using poly(A)⁺ αT3-1 RNA was performed as described (Sambrook et al., 1989, Molecular Cloning: A Laboratory Manual. Cold Spring Harbor
35 Laboratory Press, Cold Spring Harbor, NY). In situ

hybridization using ^{35}S -UTP labeled cRNA was performed on free-floating pituitary sections following published methods (Gall & Isackson, 1989, Science 245: 758-761). Sections were mounted and exposed to
5 Amersham Beta-max film for 3 days or dipped in radioactive emulsion and developed after 17 days.

6.2. RESULTS

6.2.1. cDNA CLONING OF A FUNCTIONAL MURINE GnRH-R

10 RNA from the mouse gonadotrope cell line, $\alpha\text{T3-1}$ [5], which directs the expression of a functional GnRH-R in *Xenopus* oocytes (Sealfon et al., 1990, Mol. Endocrinol 4: 119-124), was used to synthesize cDNA
15 for PCR with degenerate oligonucleotides corresponding to conserved motifs of the G protein-coupled receptors (GPRs; see Probst et al., 1992, DNA and Cell Biol. 11: 1-20). PCR products were subcloned and sequenced, and antisense oligomers synthesized for a hybrid-arrest
20 assay (Kawashi, 1985, Nuc. Acids Res. 13: 4991-5004). An oligonucleotide corresponding to clone WZ7, when co-injected with $\alpha\text{T3-1}$ and rat brain RNA, completely abolished the expression of the GnRH-R in oocytes but did not affect expression of the brain $5\text{HT}_{1\text{C}}$ receptor
25 (Fig. 1). A second antisense oligonucleotide, representing a different segment of WZ7, also completely and specifically eliminated GnRH-R expression in all oocytes tested (n=16). Clone WZ7 was used as a probe to screen an $\alpha\text{T3-1}$ bacteriophage
30 cDNA library and seven positive plaques were purified. To test whether the clone with the largest insert of 1.3 kb, WZ25, encodes a functional GnRH-R, it was subcloned for RNA synthesis and oocyte expression. All synthetic RNA-injected oocytes (n>50), when
35 exposed to GnRH, demonstrated a large depolarizing response characteristic of GnRH-R expression (Fig. 2).

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The reversal potential (V_r) and calcium-dependence of the response to GnRH induced in oocytes by WZ25 RNA transcript were similar to those previously obtained using α T3-1 RNA (Sealfon et al., 1990, Mol. Endocrinol 4: 119-124). The V_r of the current elicited by GnRH was -27 ± 0.79 mV ($n=7$), consistent with that of the chloride ion in oocytes (Barish, 1983, J. Physiol. 342: 309-325). The GnRH-elicited response was completely abolished by preloading the oocyte with 5 mM EGTA one hour before recording ($n=4$), but was not significantly affected by the absence of Ca^{2+} in the perfusate ($n=7$). Thus the receptor expressed from clone WZ25 exhibited a response mediated through the activation of the oocyte's calcium-dependent chloride current by intracellular calcium, as is characteristic of receptors that cause phosphatidylinositol hydrolysis (see Dascal, 1987, CRC Crit. Rev. Biochem. 417: 47-61). The pharmacology of the response obtained was in agreement with expression of the mammalian GnRH-R. The GnRH agonist [D-Ser(t-Bu)⁶, Pro⁹-NHET]GnRH (100 nM buserelin, $n=6$) elicited a depolarizing current in RNA-injected oocytes. In the presence of equimolar weak GnRH antagonist [D-Phe^{2,6}, Pro³]GnRH, there was a 60% reduction in the response to GnRH, in comparison with the response to GnRH alone (1880 ± 551 nA, $n=5$, and 4756 ± 1082 nA, $n=4$, respectively). Two potent GnRH antagonists completely eliminated the GnRH-elicited current (Fig. 2A).

To further characterize the receptor encoded by this cDNA clone, radioligand binding assays were performed on membranes purified from oocytes injected with the WZ25 RNA transcript. The GnRH agonist [D-Ala⁶, N α Me-Leu⁷, Pro⁹-NHET]GnRH (GnRH-A) bound with high affinity to membranes of oocytes injected with synthetic RNA (Fig. 2B). Displacement of ¹²⁵I-GnRH-A by

GnRH-A revealed similar Kds of 4.5 and 2.9 nM in WZ25 RNA-injected oocyte membranes and α T3-1 cell membranes respectively. Displacement by GnRH of GnRH-A bound to the cloned receptor was an order of magnitude less effective, as has been previously reported for α T3-1 membranes (Horn et al., 1991, Mol. Endocrinol. 5: 347-355). Thus the hybrid-arrest and expression data confirm that clone WZ25 represents the mouse GnRH-R.

10 6.2.2. CHARACTERIZATION OF THE CODING
 SEQUENCE OF MURINE GnRH-R

 The nucleotide (SEQ. ID NO: 1) and corresponding predicted amino acid sequence (SEQ. ID NO: 2) of clone WZ25 are shown in Figure 3. The longest open reading
15 frame encodes a 327 amino acid protein (relative molecular mass, $M_r=37,683$). The larger size reported for the binding subunit of the solubilized rat GnRH-R, M_r 50,000-60,000 (Hazum et al., 1986, J. Biol. Chem. 261: 13043-13048; Iwashita et al., 1988, J. Mol. Endocrinol. 1: 187-196), may be due to receptor
20 glycosylation. Three consensus N-linked glycosylation sites are present, two in the N-terminus and one in the putative first extracellular loop. The first ATG is believed to represent the translation initiation
25 site because it closely approximates a Kozak consensus sequence (Kozak, 1987, Nuc. Acids Res. 15: 8125-8148) and a second cDNA clone with additional 5' sequence contains two nonsense codons in this reading frame at positions -54 and -57. Thus translation initiating at
30 any upstream start sites would terminate before reaching the correct open reading frame. There is no polyadenylation signal and the apparent poly(A) tail most likely represents oligo(dT) priming in the 3'-untranslated region during library construction. The
35 functional GnRH-R cDNA isolated is 1.3 kb whereas the mRNA containing this sequence is approximately 4.6 kb

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as determined by sucrose gradient (Sealfon et al., 1990, Mol. Endocrinol 4: 119-124) and northern blot analysis (Fig. 5B). PCR analysis of 40 positive plaques identified by primary library screening suggests that the GnRH-R mRNA contains both additional 5'- and additional 3'-untranslated sequence.

Hydrophobicity analysis of the deduced protein demonstrates seven stretches of highly hydrophobic amino acids with 20-30% sequence similarity to other GPRs with the highest degree of homology to the interleukin-8 receptor (Fig. 4). While several highly conserved residues are noted in the GnRH-R, such as the cysteines present in each of the first two extracellular loops which stabilize many receptors, several features of the GnRH-R are unusual. For example, the highly conserved transmembrane II aspartate/glutamate, which has been found to be essential for the function of many GPRs, is replaced by an asparagine. The GnRH-R is nearly the smallest member of the GPR superfamily and, unlike any other GPR, it lacks a polar cytoplasmic C-terminus. The putative first cytoplasmic loop is longer than any other GPR. Unique among GPRs, the GnRH-R may activate via dimerization (Conn et al., 1982, Nature 296: 653-655; Gregory & Taylor, 1982, Nature 300: 269-271). Its unusual structure may subserve this proposed mechanism of activation.

Another deviation from other GPRs is the substitution of serine for the conserved tyrosine located adjacent to transmembrane III. This creates a potential phosphorylation site, unique to the GnRH-R, in a domain critical for signal transduction of other GPRs. Phosphorylation of the C-terminus, which is absent in the GnRH-R, contributes to desensitization of several GPRs (see Probst et al., 1992, DNA and Cell

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Biology 11: 1-20). It will be interesting to determine whether the novel phosphorylation site of the GnRH-R mediates receptor desensitization. Other potential regulatory phosphorylation sites are also present (Fig. 3).

The presence of GnRH-R mRNA in a variety of neuroendocrine cell lines was studied by solution hybridization/nuclease protection assay (Fig. 5A). GnRH-R mRNA was detected in α T3-1 cells and in mouse pituitary, but not in GnRH neuron-derived (GT-1), corticotroph (AtT20) or somatolactotroph (GH3) cell lines at the limits of detection of the assay. The absence of detectable GnRH-R mRNA in the GT-1 and AtT-20 cell lines has been confirmed using higher concentrations of RNA in the solution-hybridization/nuclease protection assay (Dr. Andrea C. Gore, unpublished data). Figure 5C shows the distribution of the GnRH-R mRNA in rat anterior pituitary. Labelling was heterogeneously distributed throughout the gland, a pattern previously observed by GnRH-R autoradiography (Badr & Pelletier, 1988, *Neuropeptides* 11: 7-11). Bright-field and dark-field microscopy reveals clustering of the cells expressing the GnRH-R mRNA (Fig. 5 E,F).

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7. EXAMPLE: CLONING AND CHARACTERIZATION OF HUMAN GnRH-R

The subsections below describe the cloning of complementary DNA representing the human GnRH-R and confirms its identity using *Xenopus* oocyte expression. In addition, the human GnRH-R was expressed in COS-1 cells and was shown to be functionally active.

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7.1. MATERIALS AND METHODS

7.1.1. CLONING OF HUMAN GnRH-R

1.2 million plaques of a GT10 human pituitary cDNA library (Clontech) were probed at high stringency (Sambrook et al., 1989 Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.) with the mouse GnRH-R insert (Tsutsumi et al., 1992 Mol. Endocrinol. 6:1163-1169) which had been ³²P-labeled via random hexamer priming. Thirty-two positive plaques were identified on duplicate filters; ten were selected for further characterization and six successfully purified through subsequent screening. The clone with the largest insert was subcloned into the EcoRI site of pBluescript II SK⁺ (construct LC27-4) and both strands repeatedly sequenced on an Applied Biosystems automated sequencer (Foster City, CA, USA) using synthetic oligonucleotide primers. The sequence was analyzed using the Wisconsin GCG package on a VAX computer.

7.1.2. EXPRESSION IN XENOPUS OOCYTES

Construct LC27-4 was linearized and capped RNA transcript synthesized using T3 RNA polymerase. Oocyte preparation and electrophysiology were performed as previously described (Sealfon et al., 1990, Mol. Endocrinol. 4:119-124). Cells were injected with 1-10 ng of synthetic transcript and electrophysiology recorded via two-electrode voltage clamp 48 hours later. All agonists and antagonists were applied at a concentration of 0.2 μ M. Antagonists were introduced into the bath 3 minutes prior to GnRH exposure.

The following GnRH analogs were used:

GnRH-A: [D-Ala⁶, N-Me-Leu⁷, Pro⁹-NH₂]GnRH; antagonist 5: [D-pGlu¹, D-Phe², D-Trp^{3,6}]GnRH; antagonist 6:

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[Ac-D-NaI(2)¹, D- α -Me-pCl-Phe², D-Trp³, D-Arg⁶, D-Ala¹⁰]GnRH;
antagonist 13: [Ac-D-NaI¹, D- α -4-ClPhe², D-Pal³, D-Arg⁶,
D-Ala¹⁰]GnRH; antagonist 27: [Ac-D-NaI(2)¹, D- α -Me-pCl-
Phe², D-Trp³, N- ϵ -ipr-Lys⁵, D-Tyr⁶, D-Ala¹⁰]GnRH (Van der Spuy
5 et al., 1987, in LHRH and its Analogs: Contraceptive
and Therapeutic Applications (Vickery, B.H. and
Nestor, J.J. eds) NTP Press, Lancaster. Buserelin [D-
Ser(But)⁶, Pro⁹]GnRH) was a generous gift of Hoechst-
Roussel Pharmaceuticals (Somerville, NJ, USA).

10

7.1.3. TRANSFECTION OF COS-1 CELLS

The human GnRH-R cDNA was subcloned into an
expression vector, pSV2A, containing an SV40 early
promoter. COS-1 cells were transiently transfected
15 with the pSV2A-human Gn-RHR construct using the DEAE-
dextran method (Keown et al., 1990, in Methods in
Enzymology, VI. 185 (Goeddel, D.V., ed.) pp. 527-537,
Academic Press, New York). In studies on GnRH
binding, 3 x 10⁶ cells/10 cm dish were transfected with
20 15 μ g DNA. For studies on inositol phosphate
production, 1.8 x 10⁵ cells/well (12-well plates) were
transfected with 1.5 μ g DNA. Cells were assayed 48
hours after transfection.

25

7.1.4. RECEPTOR BINDING

Cell membranes were prepared from transfected
cells with a single centrifugation step as described
for rat pituitaries (Millar et al., 1989, J. Biol.
Chem. 264:21007-21013). The receptor binding assay
30 was performed as previously described (Tsutsumi et
al., 1992, Mol. Endocrinol. 6:1163-1169) using ¹²⁵I-
GnRH-A. 10⁻⁷ M GnRH-A was used to estimated non-
specific binding.

35

7.1.5. STIMULATION OF INOSITOL PHOSPHATE PRODUCTION

GnRH-stimulated inositol phosphate (IP) production was determined as described (Davidson et al., 1990, Endocrinology 126:80-87). Accumulation of [³H]IP in the presence of LiCl was used as an index of inositol phosphate turnover. Briefly, transfected cells were labelled overnight with [³H]inositol and stimulated with 1.0 μ M GnRH in the presence of LiCl. The reaction was terminated by the addition of a perchloric acid solution and phytic acid. After neutralizing with KOH, the inositol phosphates were separated on a Dowex ion exchange column and counted.

7.1.6. NORTHERN BLOT AND PCR ANALYSIS

RNA was prepared from six human pituitaries (five male, one female, age 30-45) and human testis (age 80) by extraction with guanidinium thiocyanate followed by centrifugation in cesium chloride (Sambrook et al., 1989 in Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY). Pituitary (1.6 μ g) and testis (0.9 μ g) poly(A) RNA prepared using the Promega PolyA Tract mRNA isolation system was electrophoresed through a 1% agarose, 2.2 M formaldehyde gel, transferred to nitrocellulose membrane (HYbond-C extra, Amersham) in 20 x SSC, and fixed under vacuum at 80°C. The insert from construct LC27-4 was labelled to a specific activity of 7.2×10^8 cpm/ μ g using Amersham Megaprime Labelling Kit. Blots were prehybridized (2 h) and hybridized (overnight) in 2 x Pipes, 50% formamide, 0.5% SDS, 100 μ g/ml herring sperm DNA at 42°C, followed by washing (final wash 0.2 x SSC, 0.1% SDS 60°C for 10 minutes). In order to delineate the extent of 5'- and 3'-untranslated sequence in the

- 40 -

human RNA, the clones identified on duplicate filters in the primary library screening which were not purified were used as PCR templates with pairs of primers directed against the GT10 cloning site and the known human GnRHR insert. The PCR reaction products obtained were compared with those obtained using clone LC27-4 as the PCR template on 1% agarose gels.

7.2 RESULTS

7.2.1 CLONING AND CHARACTERIZATION OF HUMAN GNRH-R

Sequencing of clone LC27-4 identified a 2160 bp insert (Fig. 9). The largest open reading frame (1008 bp) extends to the 5'-end of the clone. The translation initiation site is assigned to the first ATG in part because of the presence of a Kozak consensus sequence (Kozak, 1987 Nucleic Acids Res. 15:8125-8148). Because the clone characterized remains in reading frame in its entire 5'-extent, the existence of additional upstream initiation sites cannot be excluded. However, the presence of additional 5'- coding region is considered unlikely because of the high homology with the mouse receptor of which the translation initiation site can be assigned with greater certainty (Tsutsumi et al., 1992). The human receptor cDNA thus contains a 984 bp reading frame which encodes a 328 amino acid protein with 90% identity to the predicted sequence of the mouse receptor. The long 3'-untranslated region contains no polyadenylation signal.

Northern blot analysis was performed to determine the size of the full length human GnRHR RNA. The probe revealed a single band of ~4.7 kb in human pituitary poly(A) RNA (Fig. 10). No signal was detected in poly(A) RNA purified from human testis or

with a human β -actin cDNA control. To determine the extent of the 5'- and 3'- untranslated domains of the RNA, PCR analysis of the phage isolates from the primary library screening was undertaken. An
5 antisense oligonucleotide primer representing sequence near the 5'-end of the LC27-4 insert or a sense primer near the 3'-end of the same sequence was used in conjunction with primers designed against the adjacent GT1-cloning site to map the unpurified clones. The
10 longest PCR products identified had 1.3 of additional 5'-sequence and 0.3 kb of additional 3'-sequence (not shown). These data suggest that the GnRHR mRNA contains at least 1.3kb of 5'-untranslated sequence and 1.5 kb of 3'-untranslated sequence. Based on the
15 Northern blot data, this suggests that additional untranslated sequence (<1 kb) is not contained in any of the clones isolated.

Hydrophobicity analysis (Kyte-Doolittle) identified the seven hydrophobic domains
20 characteristic of G--protein coupled receptors (see Fig. 9). As was found of the predicted structure of the mouse receptor, the human GnRHR lacks essentially any C-terminal intracellular domain. Two potential N-linked glycosylation sites are present, one in each
25 of the first two extracellular domains. Several cytoplasmic serine and threonine residues are found on intracellular domains and may serve as regulatory phosphorylation sites.

30 7.2.2 XENOPUS OOCYTE INJECTIONS

The largest clone isolated, LC27-4, contained a -2.2 kb insert. To test whether this clone encoded a functional human GnRHR, synthetic RNA transcript was injected into Xenopus oocytes. All RNA-injected
35 oocytes developed large depolarizing currents upon

exposure to 2×10^{-7} M GnRH ($n = 17$) or 2×10^{-7} M buserelin ($n = 6$; Fig. 1) which were indistinguishable from the responses obtained following expression of the mammalian GnRHR in oocytes using tissue or cell
5 line RNA (Sealfon et al., 1990 Mol Endocrinol. 4:119-124). These responses were completely blocked by equimolar concentrations of two potent GnRH receptor antagonists ($n = 5$ for each; Fig. 6).

10 7.2.3 EXPRESSION OF HUMAN GnRH-R IN COS-1 CELLS

To further characterize the cloned human GnRHR, the receptor was expressed in COS-1 cells. Binding data using membranes from COS-1 cells transfected with the human GnRHR construct are presented in Figure 7.

15 The displacement of GnRH-A by GnRH-A, GnRH and antagonist 5 had dissociation constants of 0.97 nM, 2.8 nM and 8.4 nM respectively, values similar to those previously obtained with human pituitary membranes (Wormald et al., 1985 J. Clin. Endocrinol.
20 Metab. 61:1190-1198).

The receptor expressed in COS-1 cells was functional and found to be coupled to inositolphosphate metabolism. An 8-fold increase in phosphoinositol metabolism was achieved at maximal receptor
25 stimulation and the EC_{50} of GnRH was 3nM. The stimulation of PI turnover induced by (10^{-8}) M GnRH was inhibited by a GnRH antagonist in a concentration - dependent manner (FIG. 8). GnRH-stimulated (10^{-8} M) inositol phosphate production was inhibited by
30 antagonist 13 with an IC_{50} of 6.7×10^{-9} M and by antagonist 5 with an IC_{50} of 1.05×10^{-7} M (not shown), giving K_d values of 2.1×10^{-10} M and 3.6×10^{-9} M respectively (Leslie, F.M., 1987 Pharmacol. Rev.
39:197-247).

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The present invention is not to be limited in scope by the exemplified embodiments which are intended as illustrations of single aspects of the invention, and any clones, DNA or amino acid sequences
5 which are functionally equivalent are within the scope of the invention. Indeed, various modifications of the invention in addition to those skilled in the art from the foregoing description and accompanying drawings. Such modifications are intended to fall
10 within the scope of the appended claims.

It is also to be understood that all base pair sizes given for nucleotides are approximate and are used for purposes of description.

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Sealfon, Stuart C.
- (ii) TITLE OF INVENTION: Cloning and Expression of
Gonadotropin-Releasing Hormone Receptor
- (iii) NUMBER OF SEQUENCES: 4
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Pennie & Edmonds
 - (B) STREET: 1155 Avenue of the Americas
 - (C) CITY: New York
 - (D) STATE: New York
 - (E) COUNTRY: U.S.A.
 - (F) ZIP: 10036-2711
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: To be assigned
 - (B) FILING DATE: 21-JUN-1993
 - (C) CLASSIFICATION:
- (vii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Misrock, S. Leslie
 - (B) REGISTRATION NUMBER: 18,872
 - (C) REFERENCE/DOCKET NUMBER: 6923-035
- (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 212 790-9090
 - (B) TELEFAX: 212 869-8864/9741
 - (C) TELEX: 66141 PENNIE

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1227 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: DNA (genomic)

- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 43..1023

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Ala Ser Leu Glu Gln Asp Pro Asn His Cys Ser Ala Ile Asn Asn Ser					
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310 315 320	
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Phe Ser Leu	
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(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 327 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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	20						25					30			
Val	Ser	Gly	Lys	Ile	Arg	Val	Thr	Val	Thr	Phe	Phe	Leu	Phe	Leu	Leu
	35				40							45			
Ser	Thr	Ala	Phe	Asn	Ala	Ser	Phe	Leu	Leu	Lys	Leu	Gln	Lys	Trp	Thr
	50				55						60				
Gln	Lys	Arg	Lys	Lys	Gly	Lys	Lys	Leu	Ser	Arg	Met	Lys	Val	Leu	Leu
65				70					75					80	
Lys	His	Leu	Thr	Leu	Ala	Asn	Leu	Leu	Glu	Thr	Leu	Ile	Val	Met	Pro
	85						90						95		
Leu	Asp	Gly	Met	Trp	Asn	Ile	Thr	Val	Gln	Trp	Tyr	Ala	Gly	Glu	Phe
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Ala	Phe	Met	Met	Val	Val	Ile	Ser	Leu	Asp	Arg	Ser	Leu	Ala	Ile	Thr
	130					135					140				
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145				150					155					160	
Ser	Leu	Ala	Trp	Ile	Leu	Ser	Ile	Val	Phe	Ala	Gly	Pro	Gln	Leu	Tyr
	165							170						175	
Ile	Phe	Arg	Met	Ile	Tyr	Leu	Ala	Asp	Gly	Ser	Gly	Pro	Thr	Val	Phe
	180						185						190		
Ser	Gln	Cys	Val	Thr	His	Cys	Ser	Phe	Pro	Gln	Trp	Trp	His	Gln	Ala

SUBSTITUTE SHEET

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195	200	205
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Leu His Gln Asp Pro Arg Lys Leu Gln Met	Asn Gln Ser Lys Asn Asn	
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	260 265 270	
Thr Ser Phe Val Val Cys Trp Thr Pro Tyr Tyr Val Leu Gly Ile Trp		
	275 280 285	
Tyr Trp Phe Asp Pro Glu Met Leu Asn Arg Val Ser Glu Pro Val Asn		
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(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 2160 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: DNA (genomic)

- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 25..1008

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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Asn Gln Asn His Cys Ser Ala Ile Asn Asn Ser Ile Pro Leu Met Gln	
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Gly Asn Leu Pro Thr Leu Thr Leu Ser Gly Lys Ile Arg Val Thr Val	
30 35 40	
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60 65 70	
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Ser Arg Met Lys Leu Leu Leu Lys His Leu Thr Leu Ala Asn Leu Leu	
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SUBSTITUTE SHEET

-48-

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CCC TAC TAT GTC CTA GGA ATT TGG TAT TGG TTT GAT CCT GAA ATG TTA Pro Tyr Tyr Val Leu Gly Ile Trp Tyr Trp Phe Asp Pro Glu Met Leu 285 290 295	915
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SUBSTITUTE SHEET

-49-

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CTATATAAAG GGAAATTTA GAGGGGTTAG GATTTGGACA GGGGTTTAGA ACATTCTCT      1728
AAGCTATCTA GTCTGTGAG TTTGTGGCA TTAATTGCCA TAAAATAACA TGTTTCCAA      1788
TGCAACTAAG AAAATACTCA TAGTGAGTAC GCTCTATGCA TAGTATGACT TCTATTTAAT      1848
GTGAAGAATT TTTTGTCTCT CTCCTGATCT TACTAAATCC ATATTTTATA AATGAACTGA      1908
GAATAATTAA CAAATTAAG CAAATGCACA AGCAAAAGAT GCTTGATACA CAAAAGGAAC      1968
TCTGGAGAGA AAACACAGC TTCAGTCTGT ACAGATCAA GAAGACAGAA CATGTCAGGG      2028
GAAGGAGGAA AGATCTTGAT GCAGGGTTT TTAACCTGCA GTCTATGCAC AACACTATAT      2088
TTCCATGTAA TGTTTTTATT TCAGCCCTAT TTGTATTATT TTGTGCATT AAAAACACA      2148
ATCTTAAGGC CG                                          2160

```

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 328 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

```

Met Ala Asn Ser Ala Ser Pro Glu Gln Asn Gln Asn His Cys Ser Ala
 1           5           10           15
Ile Asn Asn Ser Ile Pro Leu Met Gln Gly Asn Leu Pro Thr Leu Thr
 20           25           30
Leu Ser Gly Lys Ile Arg Val Thr Val Thr Phe Phe Leu Phe Leu Leu
 35           40           45
Ser Ala Thr Phe Asn Ala Ser Phe Leu Leu Lys Leu Gln Lys Trp Thr
 50           55           60
Gln Lys Lys Glu Lys Gly Lys Lys Leu Ser Arg Met Lys Leu Leu Leu
 65           70           75           80
Lys His Leu Thr Leu Ala Asn Leu Leu Glu Thr Leu Ile Val Met Pro
 85           90           95
Leu Asp Gly Met Trp Asn Ile Thr Val Gln Trp Tyr Ala Gly Glu Leu
100          105          110
Leu Cys Lys Val Leu Ser Tyr Leu Lys Leu Phe Ser Met Tyr Ala Pro
115          120          125

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SUBSTITUTE SHEET

-50-

Ala Phe Met Met Val Val Ile Ser Leu Asp Arg Ser Leu Ala Ile Thr
 130 135 140
 Arg Pro Leu Ala Leu Lys Ser Asn Ser Lys Val Gly Gln Ser Met Val
 145 150 155 160
 Gly Leu Ala Trp Ile Leu Ser Ser Val Phe Ala Gly Pro Gln Leu Tyr
 165 170 175
 Ile Phe Arg Met Ile His Leu Ala Asp Ser Ser Gly Gln Thr Lys Val
 180 185 190
 Phe Ser Gln Cys Val Thr His Cys Ser Phe Ser Gln Trp Trp His Gln
 195 200 205
 Ala Phe Tyr Asn Phe Phe Thr Phe Ser Cys Leu Phe Ile Ile Pro Leu
 210 215 220
 Phe Ile Met Leu Ile Cys Asn Ala Lys Ile Ile Phe Thr Leu Thr Arg
 225 230 235 240
 Val Leu His Gln Asp Pro His Glu Leu Gln Leu Asn Gln Ser Lys Asn
 245 250 255
 Asn Ile Pro Arg Ala Arg Leu Lys Thr Leu Lys Met Thr Val Ala Phe
 260 265 270
 Ala Thr Ser Phe Thr Val Cys Trp Thr Pro Tyr Tyr Val Leu Gly Ile
 275 280 285
 Trp Tyr Trp Phe Asp Pro Glu Met Leu Asn Arg Leu Ser Asp Pro Val
 290 295 300
 Asn His Phe Phe Phe Leu Phe Ala Phe Leu Asn Pro Cys Phe Asp Pro
 305 310 315 320
 Leu Ile Tyr Gly Tyr Phe Ser Leu
 325

WHAT IS CLAIMED IS:

1. A cDNA encoding a GnRH-R.
- 5 2. A recombinant DNA vector containing a nucleotide sequence that encodes a GnRH-R.
3. A recombinant DNA vector containing a nucleotide sequence that encodes a GnRH-R fusion
10 protein.
4. A recombinant DNA vector of Claim 2 in which the GnRH-R nucleotide sequence is operatively associated with a regulatory sequence that controls
15 gene expression in a host.
5. A recombinant DNA vector of Claim 3 in which the GnRH-R fusion protein nucleotide sequence is operatively associated with a regulatory sequence that
20 controls gene expression in a host.
6. The DNA of Claim 1, 2, 3, 4 or 5 in which the GnRH-R sequence encodes the murine GnRH-R.
- 25 7. The DNA of Claim 1, 2, 3, 4 or 5 in which the GnRH-R sequence encodes the human GnRH-R.
8. The DNA of Claim 1, 2, 3, 4 or 5 which is capable of hybridizing under stringent conditions, or
30 which would be capable of hybridizing under stringent conditions but for the degeneracy of the genetic code to the murine GnRH-R DNA sequence (SEQ. ID NO: 1) of FIG. 3 or the human GnRH-R DNA sequence (SEQ. ID NO. 3) of FIG. 9.

35

- 52 -

9. An engineered host cell that contains the recombinant DNA vector of Claim 1, 2, 3, 4 or 5.

10. An engineered cell line that contains the
5 recombinant DNA expression vector of Claim 4 and expresses the GnRH-R.

11. The engineered cell line of Claim 10 which expresses the GnRH-R on the surface of the cell.
10

12. The engineered cell line of Claim 10 or 11 which expresses human GnRH-R.

13. An engineered cell line that contains the
15 recombinant DNA expression vector of Claim 5 and expresses the GnRH-R fusion protein.

14. The engineered cell line of Claim 13 that expresses the GnRH-R fusion protein on the surface of
20 the cell.

15. The engineered cell line of Claim 13 or 14 which expresses a human GnRH-R fusion protein.

25 16. A method for producing recombinant GnRH-R, comprising:
(a) culturing a host cell transformed with the recombinant DNA expression vector of Claim 4 and which expresses the GnRH-R; and
30 (b) recovering the GnRH-R gene product from the cell culture.

17. The method of Claim 16 in which the human GnRH-R is produced.
35

18. A method for producing recombinant GnRH-R fusion protein, comprising:

- 5 (a) culturing a host cell transformed with the recombinant DNA expression vector of Claim 5 and which expresses the GnRH-R fusion protein; and
- (b) recovering the GnRH-R fusion protein from the cell culture.

10 19. The method of Claim 18 in which a human GnRH-R fusion protein is produced.

20. An isolated recombinant GnRH receptor.

15 21. The isolated GnRH receptor of Claim 20 having a murine GnRH receptor amino acid sequence.

22. The isolated GnRH receptor of Claim 20 having a human GnRH receptor amino acid sequence.

20

23. A fusion protein comprising a GnRH receptor linked to a heterologous protein or peptide sequence.

24. The fusion protein of Claim 23 in which the

25 GnRH receptor moiety has a murine GnRH receptor amino acid sequence.

25. The fusion protein of Claim 23 in which the

30 GnRH receptor moiety has a human GnRH receptor amino acid sequence.

26. An oligonucleotide which encodes an antisense sequence complementary to a portion of the GnRH-R nucleotide sequence, and which inhibits

35 transcription of the GnRH-R gene in a cell.

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27. The oligonucleotide of Claim 26 which is complementary to a nucleotide sequence encoding the transmembrane region of the GnRH-R.

5 28. A monoclonal antibody which immunospecifically binds to an epitope of the GnRH-R.

29. The monoclonal antibody of Claim 28 which competitively inhibits the binding of GnRH to the
10 GnRH-R.

30. The monoclonal antibody of Claim 28 or 29 which binds to the human GnRH-R.

15 31. A method of contraception, comprising, immunizing a host species with GnRH-R.

32. A method of contraception, comprising, immunizing a host species with GnRH-R fusion protein.
20

25

30

35

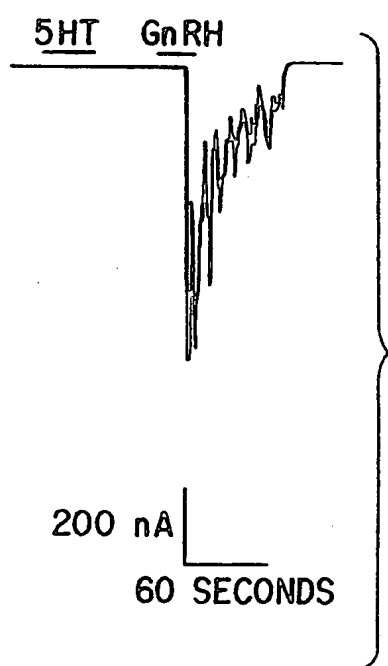


FIG. 1A

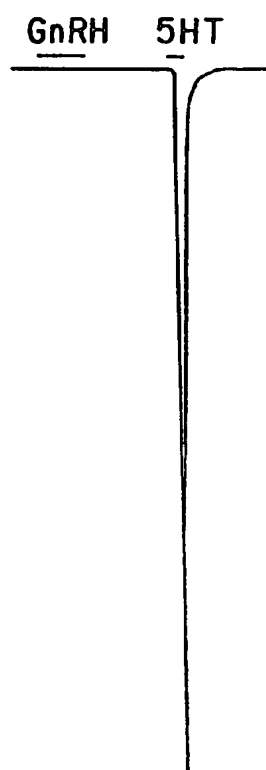


FIG. 1B

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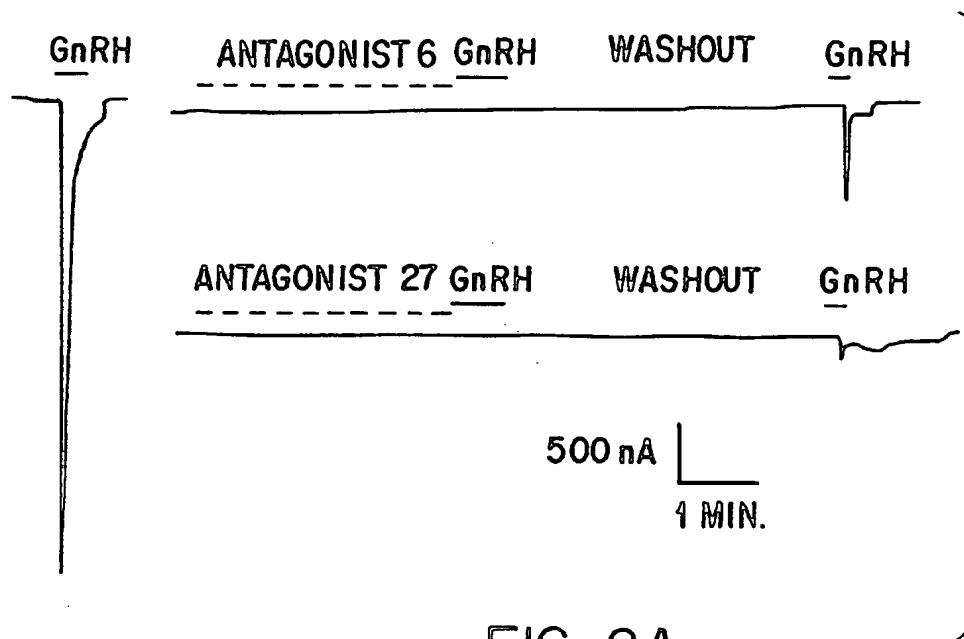


FIG. 2A

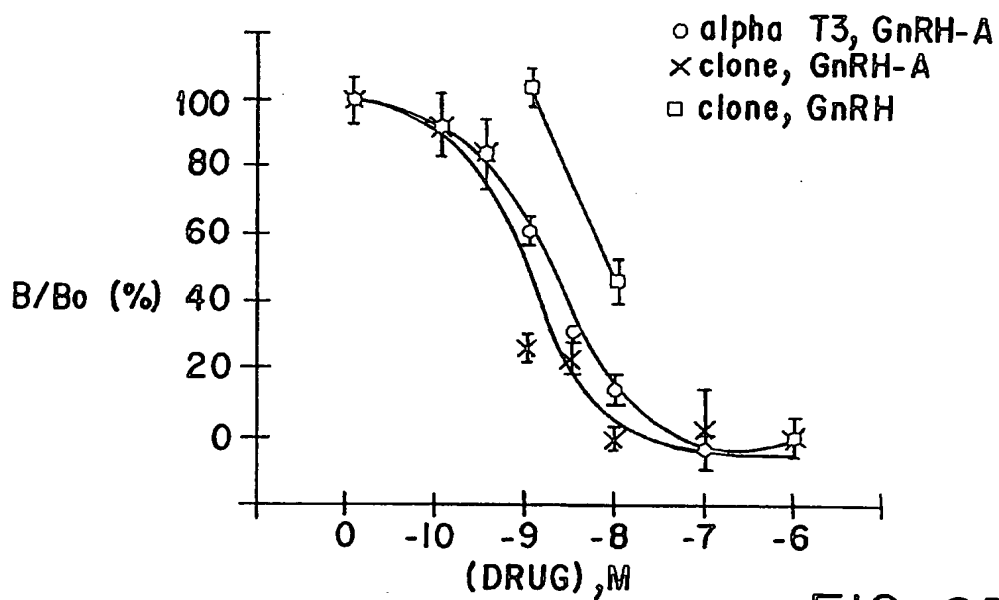


FIG. 2B

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ATGGCTAACAAATGCATCTTCTTGACGAGCACCCAAATCACTGCTCGCCATCAACAAAGC	60
M A N N A S L E Q D P N H C S A I N S	20
ATCCCCTTGATACAGGGCAAGCTCCCGACTCTAACCCTATCTGGAAAGATCCGAGTGACC	120
I P L I Q G K L P T L T V S G K I R V T	40
GTGACTTTCTTCTTTTCTACTCTCTACTGCGTTCAATGCTTCCTTCTTGTGAAGCTG	180
V T F F L F L L S T A F N A S F L L K L	60
CAGAAGTGGACTCAGAAGAGGAAGAAAGGAAAAAGCTCTCAAGGATGAAGTGCTTTTA	240
Q K W T Q K R K K G K K L S R M K V L L	80
AAGCATTTGACCTTAGCCAACTGCTGGAGACTCTGATCGTCATGCCACTGGATGGGATG	300
K H L T L A N L L E T L I V M P L D G M	100
TGGAATATTACTGTTCAAGTGGTATGCTGGGGAGTTCTCTGCAAAGTTCTCAGCTATCTG	360
W N I T V Q W Y A G E F L C K V L S Y L	120
AAGCTCTTCTCTATGTATGCCCCAGCTTTTCATGATGGTGGTGATTAGCCTGGACCGCTCC	420
K L F S M Y A P A F M M V V I S L D R S	140
CTGGCCATCACTCAGCCCCTTGCTGTACAAAGCAACAGCAAGCTTGAACAGTCTATGATC	480
L A I T Q P L A V Q S N S K L E Q S M I	160
AGCCTGGCCTGGATTCTCAGCATTGTCTTTGCAGGACCACAGTTATATATCTTCAGGATG	540
S L A W I L S I V F A G P Q L Y I F R M	180
ATCTACCTAGCAGACGGCTCTGGGCCACAGTCTTCTCGCAATGTGTGACCCACTGCAGC	600
I Y L A D G S G P T V F S Q C V T H C S	200
TTTCCACAGTGGTGGCATCAGGCCTTCTACAACTTTTTCACCTTCGGCTGCCTCTTCATC	660
F P Q W W H Q A F Y N F F T F G C L F I	220
ATCCCCCTCCTCATCATGCTAATCTGCAATGCCAAAATCATCTTTGCTCTCAGCGGAGTC	720
I P L L I M L I C N A K I I F A L T R V	240
CTTCATCAAGACCCACGCAAACTACAGATGAATCAGTCCAAGAATAATATCCCAAGAGCT	780
L H O D P R K L Q M N Q S K N N I P R A	260

FIG. 3A
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CGGCTGAGAACGCTAAAGATGACAGTCGCATTGCTACCTCCTTTGTCGTCTGCTGGACT 840
R L R T L K M T V A F A T S F V V C W T 280
*
CCCTACTATGTCCTAGGCATTGTTGTTGATCCAGAAATGTTGAACAGGGTGTC 900
P Y Y V L G I W Y W F D P E M L N R V S 300
GAGCCAGTGAATCACTTTTTCTTTCTTTGCTTTCTAAACCCGCTTCGACCCACTC 960
E P V N H F F F L F A F L N P C F D P L 320
VII
ATATATGGGTATTTCTCTTTGTAGTTGGGAGACTACACAAGAACTCAGATAGAAATAAGC 1020
I Y G Y F S L 327
TAACTAATTGCACCAATTGAGAATAAACTCAAAGCTTTTGACACACTTATATACAAGGCA 1080
GGGTTTAAGTTAGATTATCAACCTTGTTTTGTACAGAGTTTGTGTTAGAGCTTCAGA 1140
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FIG.3B

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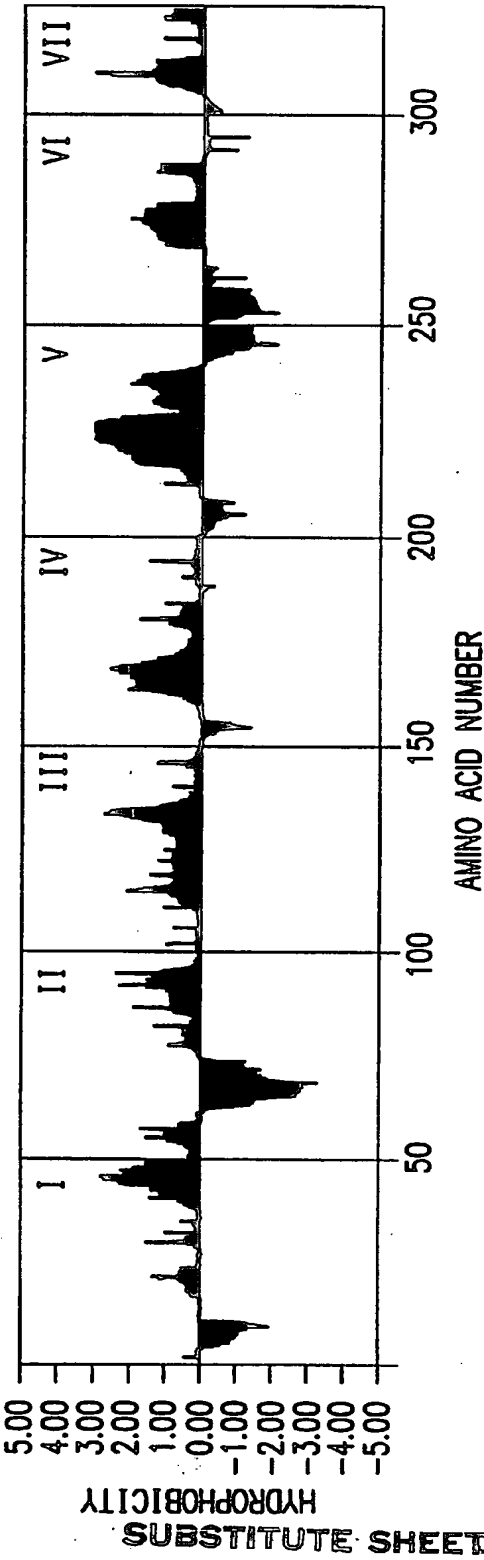


FIG. 4A

I

GnR MANVASLEQDPNHCSAINNSIPLEIGKLEPTITVSGKIRVITVTEFFLLSTAFNASFLKLQKATQKR
ILR MESDSFEDFVKGEDLSNYSYSTLPFFLLDAAPCEPESLEINKYFVMIYALVELLSLGNLSVMVILYSRVGR
SPR MDNVLPMDSDLFPNISTNTSESNQFVQPTAQIVLAAAYTVIWTSSVGNVYVVIWILAHKRMQ
β1R (9)ASEPGNLSSAAPLPDGAATAARLLVPASPPASLLPASESPEELSQQHTAGMGLLMALIVLLIVAGNVLVIVATAITPRLQ
RHO MNGTEGPNFYVPFSNATGVVRSPFEYPPQYVTAEPHQFSMLAAYMFLQIVLGFPINFLQIVYTVQHKKLQ

III

GnR KKGKKLSRMKQVLEKHLITLANLETLIVMPLDGMNITVQVYAGEFLCKVLSYKLFSMYAPAFMMVVISLDRSLATIQPLAVQSN
ILR SVTD-----MYLLNLALADLFALTPIWAASKVNGHIF--GTFCKVMSLKEVNFSGILLACISMDRYLAIVHATRLTQ
SPR TVTN-----YFLVNLAFACMAAFNTVVNF TYAVHNWVYGLFCKFHNFPIAALFASISMTAVAFDRYMAIIPHLPRLS
β1R TLTN-----LFTMSLASADVMQLLVVFPF GATIVVGRHIEYGSJFCELVTSVDVLCVTAS IETLOVIALDRYLALISPF RYQSL
RHO TPLN-----YIQNLAVADLFMVLGGF TSTLYTSLHG YFVFP TQGNLEGFATLGGELALHSLVMLATEBYVYVVKCPMSNFR

V

GnR --SKLEQSMISLAWISIVFAGPQLYIFRMIMLADGSGPTVFSQDVTH--CSFPQVWHQAFVYNFFTEGCLFTIPLLTMLIQNAKIIIFALTR
ILR -KRYLVKFICLSIVGLSLLALPVLFRRTVMSSN-----VSPACVED-MGNNTANHRMLLRILPQSFCEIVPLLIMFYGYFTLRTLFK
SPR --ATATKVVIFVIMVALLAFPGGYSTTETMPS-----RVVCMIEWPEHNRITYEKATHICVTLIYFLPLLVIAYAYTVVGITLWA
β1R LTRARAGLVCTVWATISALVSFLPILMHVWRAESD-----EARRVNDPKCCD-FVTNRATYATASSVVSFYVPLCTMAFVYL RVFREAGK
RHO -GENHAIMGVAFIMMALACAAAPDAGHSRYIPEG-----LQCSGIDYYTLKPEVNNEFVIYMEVWHTIPMIITFFQYGGQLVETVKE

FIG. 4B

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VI

GnR VLHQD-----PRKLQMNQSKNNIPARLRITLKMIVAFATSFVVCATPYMQLGIWYFDEBMLNRV
 ILR AHMGO-----KRAMRVIFAVVLIFLLCWPYNLVLLADTLMTQVIQE
 SPR SETPG-----DSSDRYHEQVSARKVKVMIVVCTFAICMLPFHVFLLPYINQDLYLKK
 β1R QVKKIDSCERRFLGGPARPPSP.(20).TAPLANGRAGRPPSRVALREKALKTLGIIMGVFTLCALPEFLANVVKAEH
 RHD -----AAQQQDESATTQKAEKEVTRMVIIMVIAELICWMPJASVAFYIFTHQGSNFGPI

VII

GnR -----SEPVNHFFLEAFINPCDDPLTYGYFSL
 ILR TCERNHIDRALDATEILGILHSELNPLIYAFIGQKFRHGLLKILAIGHLISKDSLPKDSRPSFVGSSGHTSTTL
 SPR -----FIQQVYLASMWLA MSSTMYNPJIIYCCLNDRFRLGFKHAFRCPPFI SAGDYEGLEMKSTRYLQTQSSVYKVSRLTTISTVVG(43)
 β1R ---RELVPDRLFVEENWLG YANSAPNPJIIYCRSPDFRKAFQGLLCCARRARRRHA THGDRPRASGCLARPPSPGAASDDDDDDV(43)
 RHD -----FMTIPAFFAKSAAIYNPVIYIMMNKQFRNCMLTICCGKNPLGDDEASATVSKTETSQVAPA

FIG.4C

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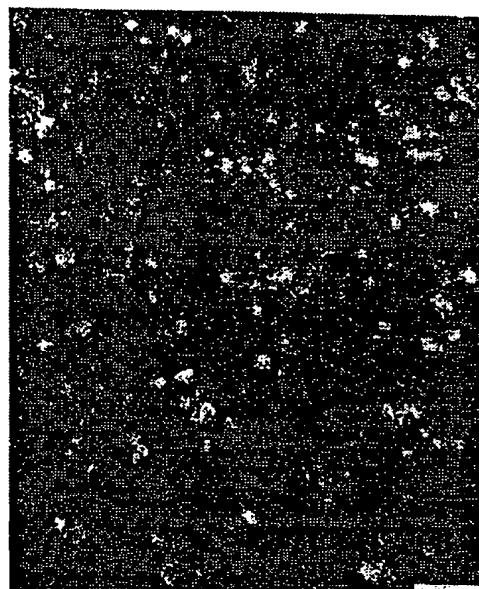
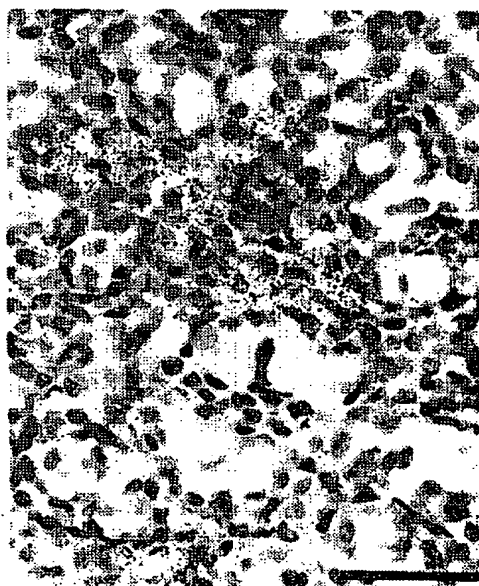
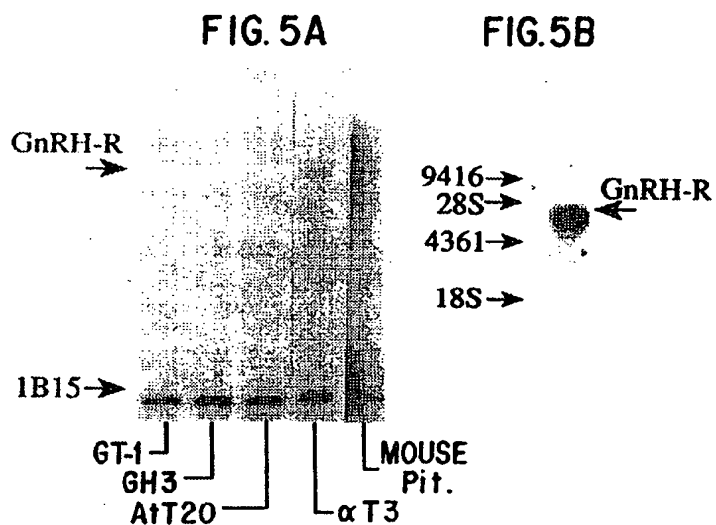


FIG. 5E

FIG. 5F

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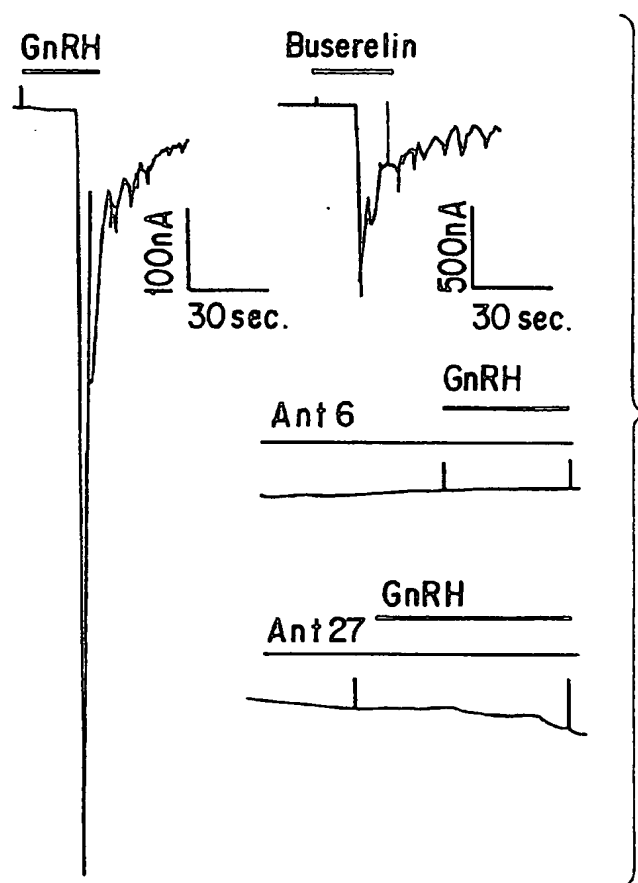


FIG. 6

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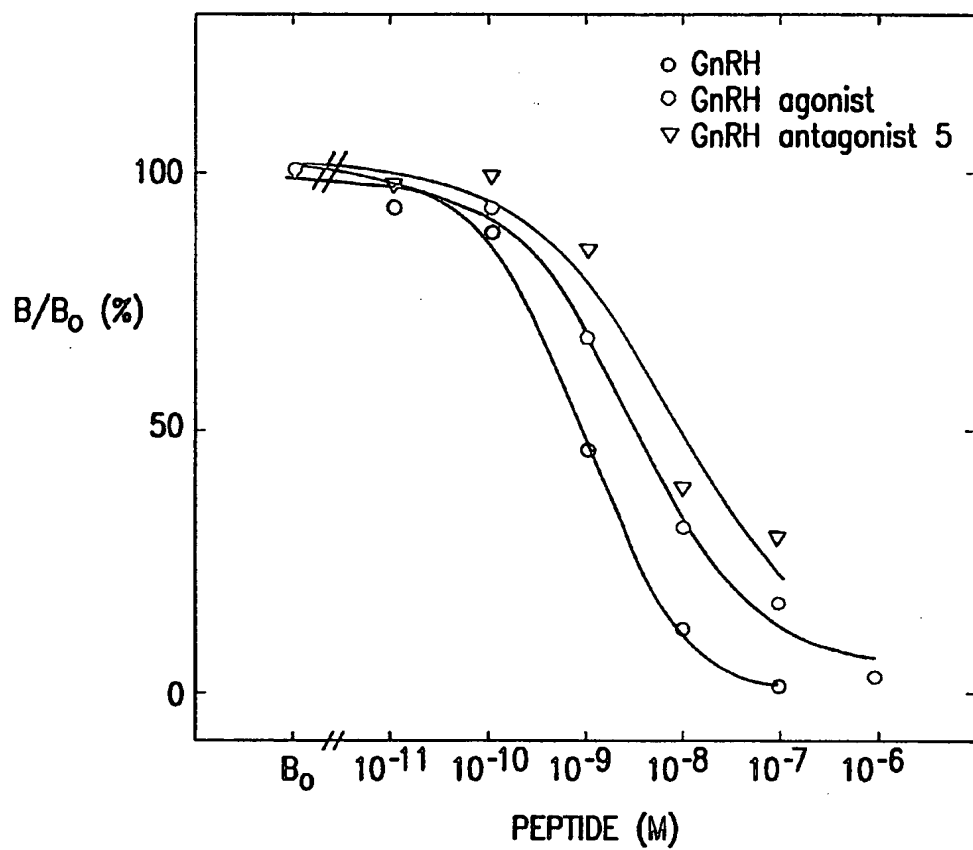


FIG.7

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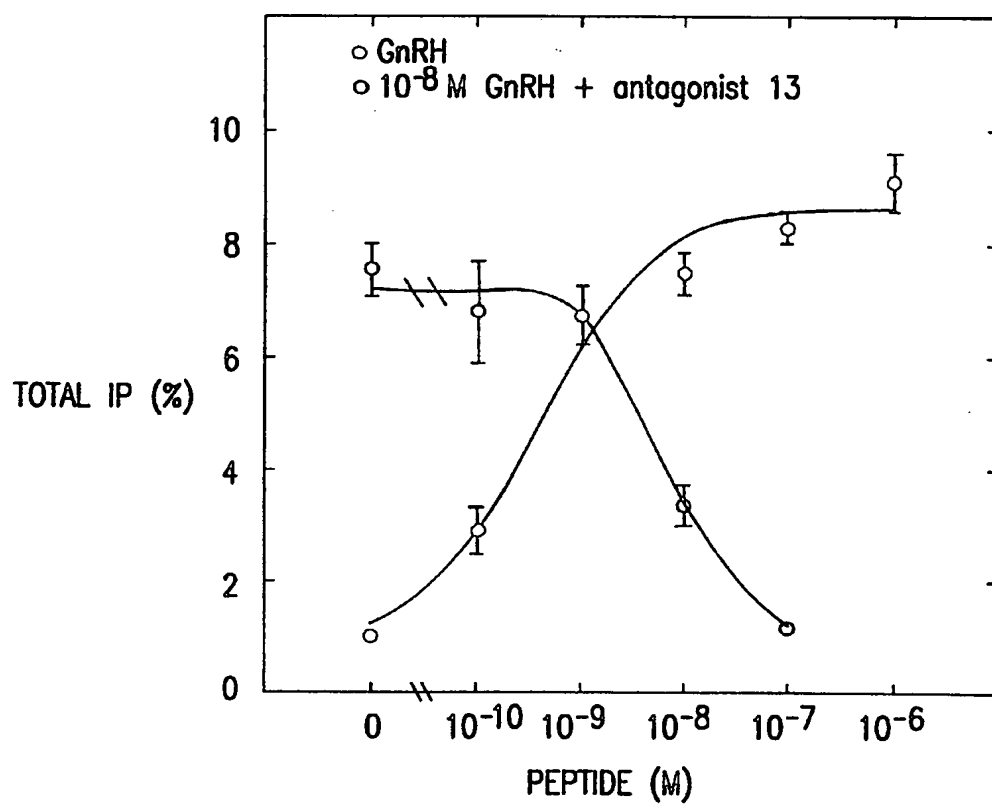


FIG.8

SUBSTITUTE SHEET

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CCGAGCCTTGTGCTCGGAAAAT -1
 ATGGCAAACAGTGCCTCTCCTGAACAGAATCAAATCACTGTTCAGCCATCAACAACAGC 60
 M A N S A S P E Q N Q N H C S A I N N S 20
 ATCCCACTGATGCAGGGCAACCTCCCACTCTGACCTTGTCTGGAAAGATCCGAGTGACG 120
 I P L M Q G N L P T L T L S G K I R V T 40
 GTTACTTTCTCTCTTTTCTGCTCTCTGCGACCTTTAATGCTTCTTTCTTGTGAAACTT 180
 V T F F L F L L S A T F N A S F L L K L 60
 CAGAAGTGGACACAGAAGAAAGAGAAAGGAAAAAGCTCTCAAGAATGAAGCTGCTCTTA 240
 Q K W T Q K K E K G K K L S R M K L L L 80
 AAACATCTGACCTTAGCCAACCTGTTGGAGACTCTGATTGTATGCCACTGGATGGGATG 300
 K H L T L A N L L E T L I V M P L D G M 100
 TGGAACATTACAGTCCAATGGTATGCTGGAGAGTTACTCTGCAAAGTTCTCAGTTATCTA 360
 W N I T V Q W Y A G E L L C K V L S Y L 120
 AAGCTTTTCTCCATGTATGCCCCAGCCTTCATGATGGTGGTGATCAGCCTGGACCGCTCC 420
 K L F S M Y A P A F M M V V I S L D R S 140
 CTGGCTATCAGAGGCCCTAGCTTTGAAAAGCAACAGCAAAGTCGGACAGTCCATGGTT 480
 L A I T R P L A L K S N S K V G Q S M V 160
 GGCCTGGCCTGGATCCTCAGTAGTGTCTTTGCAGGACCACAGTTATACATCTTCAGGATG 540
 G L A W I L S S V F A G P Q L Y I F R M 180
 ATTCATCTAGCAGACAGCTCTGGACAGACAAAAGTTTTCTCTCAATGTGTAACACACTGC 600
 I H L A D S S G Q T K V F S Q C V T H C 200
 AGTTTTTACAATGGTGGCATCAAGCATTTTATAACTTTTACCTTCAGCTGCCCTCTTC 660
 S F S Q W W H Q A F Y N F F T F S C L F 220
 ATCATCCCTCTTTTCATCATGCTGATCTGCAATGCAAAAATCATCTTCACCCTGACACGG 720
 I I P L F I M L I C N A K I I F T L T R 240
 GTCCTTCATCAGGACCCCCACGAACTACAACTGAATCAGTCCAAGAACAATATACCAAGA 780
 V L H Q D P H E L Q L N Q S K N N I P R 260
 GCACGGCTGAAGACTCTAAAAATGACGGTTGCATTGGCACTTCATTTACTGTCTGCTGG 840
 A R L K T L K M T V A F A T S F T V C W 280
 ACTCCCTACTATGTCTAGGAATTTGGTATTGGTTTGATCCTGAAATGTTAAACAGGTTG 900
 T P Y Y V L G I W Y W F D P E M L N R L 300
 TCAGACCCAGTAAATCACTTCTTCTTTCTTTGCCTTTTAAACCCATGCTTTGATCCA 960
 S D P V N H F F F L F A F L N P C F D P 320
 CTTATCTATGGATATTTTCTCTGTGATTGATAGACTACACAAGAAGTCATATGAAGAAG 1020
 L I Y G Y F S L * 328
 GGTAAGGTAATGAATCTCTCCATCTGGGAATGATTAACACAAATGTTGGAGCATGTTTAC 1080
 ATACAAACAAAGTAGGATTTACACTTAAGTTATCATTCTTTAGAACTCAGTCTTCAGA 1140
 GCCTCAATTATTAAGGAAAAGTCTTCAGGAAAAATACTAAATATTTTCTCTCCTCATA 1200
 AGCTTCTAAATTAATCTCTGCCCTTTCTGACCTCATATAACACATTATGTAGGTTTCTTA 1260

FIG.9A

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TCAC TTTCTCTTTCG CATAATAATGTACTAATATTTAAATACCTTCAGCCTAAGGCACAA	1320
GGATGCCAAAAAACAAGGTGAGAACCACAACACAGGTCTAACTCAGCATGCTTGGT	1380
GAGTTTTCTCCAAAGGGGCATATTAGCAATTAGAGTTGTATGCTATATAATACATAGAG	1440
CACAGAGCCCTTTGCCCATATAATCAACTTCCCTCTATAGTAAAAAGAAAAAAAT	1500
GAATCTATTTTCTCTTTGGCTTCAAAGCATTCTGACATTGGAGGAGTCAGTAACCAA	1560
TCCACCAACCCTCCAGCAACCTGACAAGACTATGAGTAGTTCTCCTTCATCTATTTA	1620
TGTGGTACAGGTTGTGAAGTATCTCTATATAAGGCAAATTTAGAGGGTTAGGATTTG	1680
GACAGGGGTTTAGAACATTCCTCTAAGCTATCTAGTCTGTGGAGTTGTGCAATTAATT	1740
GCCATAAAATAACATGTTTCCAAATGCAACTAAGAAAATACTCATAGTACGCTCTA	1800
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ATCCATATTTATAAATGAAGTGAATAATTAACAAAATTAAGCAAATGCACAAGCAAA	1920
AGATGCTTGATACACAAAAGGAAGTCTGGAGAGAAAAGTACAGCTTCAGTCTGTACAGAT	1980
CAAAGAAGACAGAACATGTCAGGGGAAGGAGGAAGATCTTGATGCAGGGTTCTTAACC	2040
TGCAGTCTATGCACAACACTATATTTCCATGTAATGTTTTATTTAGCCCTATTGTAT	2100
TATTTTGTGCATTTAAAAACACAATCTTAAGGCCG	2136

FIG.9B

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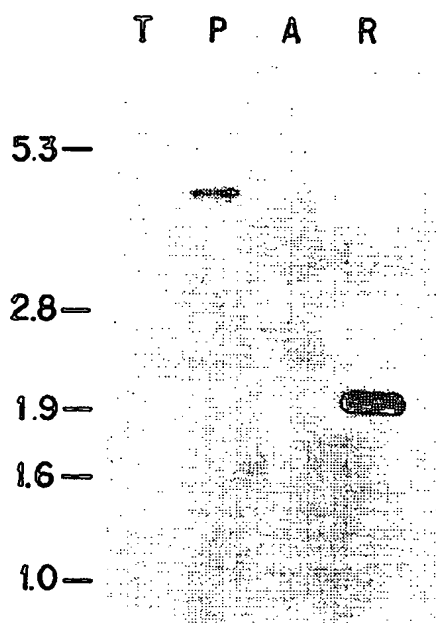
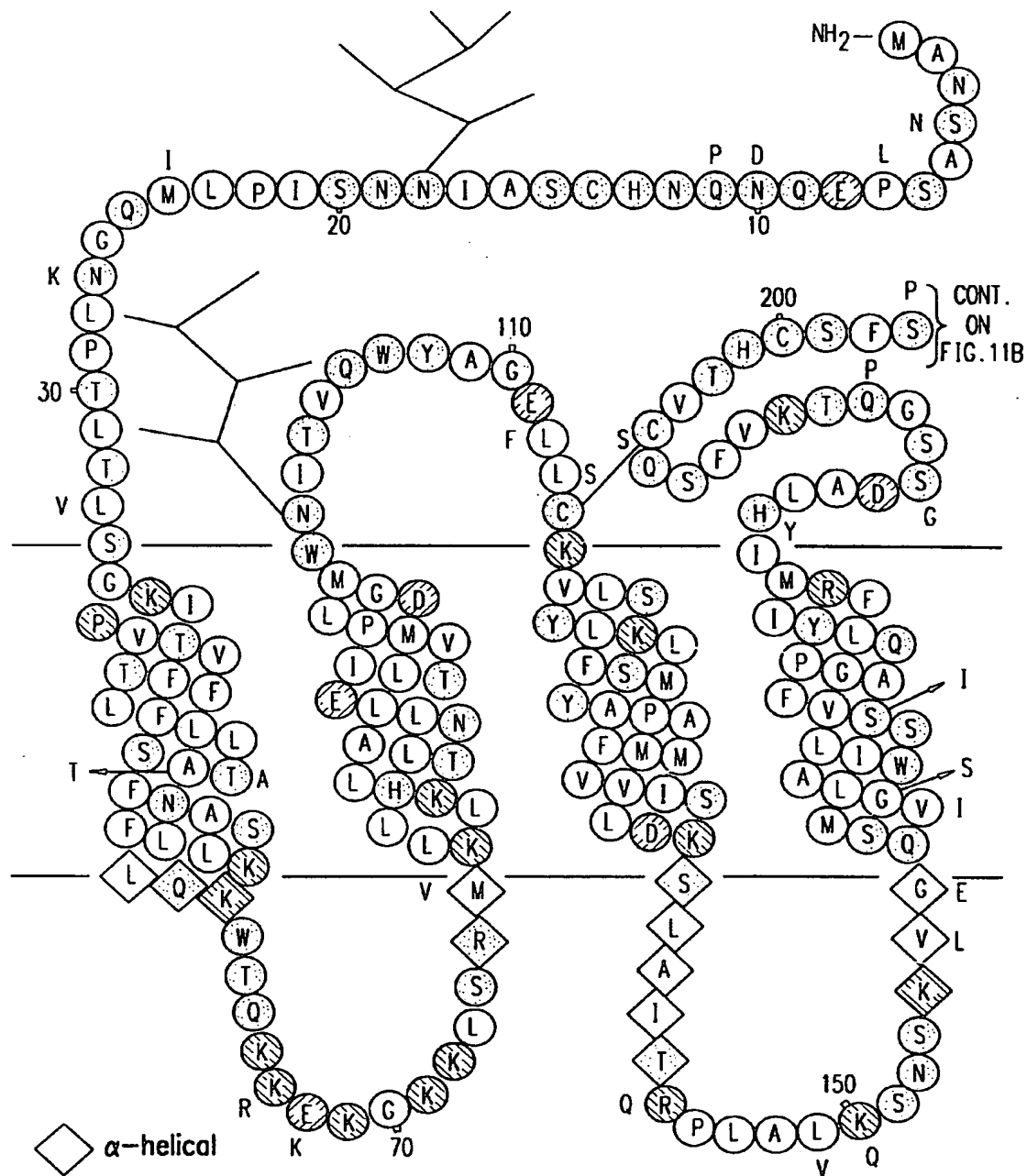
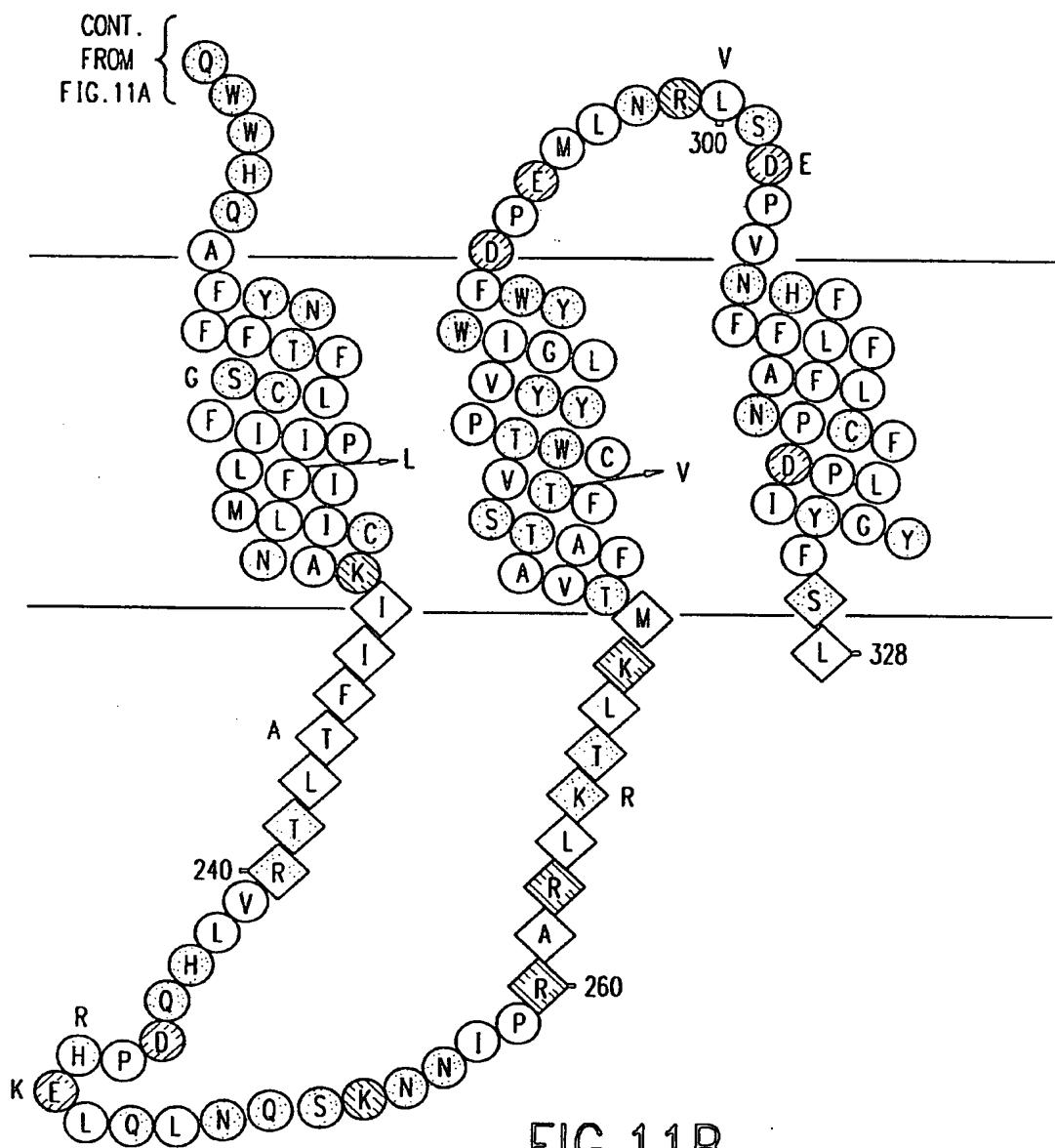


FIG. 10

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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US93/05965

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) :Please See Extra Sheet.

US CL :Please See Extra Sheet.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/91, 69.1, 69.4, 172.1; 530/387.9, 399; 536/23.5, 23.51; 514/12, 841; 424/88; 935/9

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, DIALOG

search terms: gnrh receptor

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X Y	Molecular Endocrinology, Volume 4, No. 1, issued 1990, S. Sealfon et al., "Gonadotropin-releasing hormone receptor expression in Xenopus oocytes", pages 119-124, entire document.	1-25 26-32
Y,P	US, A, 5,190,931 (INOUE ET AL) 02 MARCH 1993, entire document.	26, 27
Y	Nature, Volume 350, issued 04 April 1991, M.L. Riordan et al., "Oligonucleotide-based therapeutics", pages 442-443, entire document.	26,27
Y	Oncogene, Volume 1, issued 1987, O. Shohat et al., "Inhibition of cell growth mediated by plasmids encoding p53 anti-sense", pages 277-283, entire document.	26, 27
A	Molecular Endocrinology, Volume 6, No. 7, issued 1992, M. Tsutsumi et al., "Cloning and functional expression of a mouse	1-32

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be part of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier document published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&" document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means	
"P" document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

26 July 1993

Date of mailing of the international search report

AUG 04 1993

Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
Box PCT
Washington, D.C. 20231

Authorized officer

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Facsimile No. NOT APPLICABLE

Telephone No. (703) 308-0196

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US93/05965

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A,P	Biochem. Biophys. Res. Commun., Volume 189(1), issued 30 November 1992, S.S. Kakar et al., "Cloning, sequencing and expression of human gonadotropin releasing hormone (GnRH) Receptor", pages 289-295, entire document.	1-32
A	Mol. Cell. Endocrinol., Volume 90, issued 1992, K.A. Eidne et al., "Molecular cloning and characterisation of the rat pituitary gonadotropin-releasing hormone (GnRH) receptor", pages R5-R9, entire document.	1-32

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US93/05965

A. CLASSIFICATION OF SUBJECT MATTER:
IPC (5):

C12P 19/34, 21/06; C12N 15/00; A61K 35/14, 37/00, 39/00; C07K 13/00; C07H 21/04

A. CLASSIFICATION OF SUBJECT MATTER:
US CL :

435/91, 69.1, 69.4, 172.1; 530/387.9, 399; 536/23.5, 23.51; 514/12, 841; 424/88; 935/9